ALLEVIATING ADVERSE EFFECTS OF HEAT STRESS BY USING ORGANIC SELENIUM AND CHROMIUM FOR LOCAL LAYING HENS.

El-Samra H.A. Abo-Egla*; Z.M. Kalaba*; A.A.H. Tolba** and M.A.I. El-Deeb**

ABSTRACT

This experiment was conducted to evaluate the effect of selenium (Sel-Plex) and chromium picolinate on some blood parameter of local laying hens reared under high environmental temperatures. Two hundred and seventy laying hens (24 week-old) were divided into nine groups, 30 hens per group using two levels of selenium (0.3 and 0.5 mg/kg diet) and two levels of chromium picolinate (1200 and 1400 μg/kg diet). The control group was fed a basal diet. The obtained results can be summarized as follows: Laying hens fed the Se and Cr-supplemented diets achieved significantly higher total counts of blood erythrocytes and leukocytes, and lymphocytes, hemoglobin, packed cell volume as compared to their control counterparts. However, there were significant decreases (P≤0.05) in total monocytes but total heterophil were not affected compared with the un-supplemented control group. Dietary supplementation with Se and Cr caused significant (P≤0.05) increase in blood plasma levels of total protein, globulin, chloride, calcium, phosphorus, triiodothyronine, and thyroxine, while levels of albumin, albumin: globulin ratio, creatinine and urea were decreased as compared to those of the control group. Hens fed the Se and Cr-supplemented diets attained significantly lower (P≤0.05) levels of plasma concentrations of cholesterol, glucose, total lipids and triglycerides as well as activity of transaminases (ALT and AST) as compared to their control counterparts. From the present results, it is concluded that enhancing hen’s diets with selenium (as Sel-Plex) and chromium picolinate singly or in combination at levels of 0.3 mg Se plus 1400 μg Cr/kg diet can consider as an effective management practice for reducing the adverse effects of heat stress for laying hens.

Keywords: heat stress, local laying hens, blood parameters.

INTRODUCTION

High environmental temperature is a major problem faced by poultry farmers particularly laying hens, during summer months. High ambient temperature reduces feed intake, live weight gain, egg production, egg quality and feed efficiency (Donkoh, 1989; Siegel, 1995), thus negatively influencing the performance of poultry. Plasma corticosterone concentration also increases during heat stress (Hurwitz et al. 1980). In addition, Donkoh (1989) reported that reduced plasma protein and markedly increased blood glucose concentrations during heat stress. Such ambient temperatures decrease serum vitamin and mineral concentrations in poultry as well as humans (Ensminger et al., 1990; Anderson, 1994). Heat stress has also been shown to increase mineral excretion (Siegel, 1995). Several methods are available to alleviate the effect of high environmental temperature on poultry
performance. Because it is expensive to cool animal buildings, such methods focus mainly on manipulating the diet.

Chromium is used in the poultry diet because of the reported benefits of Chromium supplementation to laying hens (Sahin et al., 2001; Sahin et al., 2002) during cold and heat stress and because chromium is reduced during environmental stress. The primary role of Chromium in metabolism is to potentiate the action of insulin through its presence in an organometallic molecule, the glucose tolerance factor (GTF) (Anderson, 1994; Sahin et al., 2001). As Insulin metabolism influences lipid peroxidation (Gallaher et al., 1993), chromium, as an insulin potentiator, is therefore postulated to function as an antioxidant (Preuss, et al., 1997). Moreover, Chromium is thought to be essential for activating certain enzymes and for stabilizing proteins and nucleic acids (Anderson; 1994; Linder, 1991).

Selenium supplementation as a single agent or in combination has been suggested to improve performance and immune response to diseases and also to decrease the economic losses related to high temperature (Smart et al., 1995). Sodium selenite is considered the traditional source of supplementation (Leeson and Summers, 1991). Recently, organic selenium from yeast (Sel-Plex, Alltech Inc.) is more active form of selenium in chickens than selenite (Collins et al., 1993). Leeson and Summers (1991) have shown that selenium is required for maximum performance of chickens. Mahmoud and Edens (2003) found that chickens fed organic selenium as Sel-Plex, a selenized yeast, had elevated GPX activity in both blood and liver in a thermoneutral environment and after heat distress. Therefore, the present study was undertaken to evaluate the effect of dietary supplementation with organic selenium and chromium on some blood parameters of heat-stressed laying hens.

**MATERIALS AND METHODS**

**Birds and experimental design**

The present experiment was carried at Sakha Animal Production Research Station, Animal Production Research Institute, Agricultural Research Center, Ministry of Agriculture, Kafr El-Sheikh Governorate, Egypt. A total of 270 local 24-week-old laying hens (Inshas) were used in this study. Hens were randomly distributed into nine groups, each containing 30 hens in 3 replicates of 10 hens each. Laying houses were provided with 17 h light per day. The hens were randomly assigned according to initial body weights. Feed and water were given *ad libitum*. The hens were vaccinated against Marek and Newcastle diseases. Similar management conditions were maintained for all groups. The experiment was carried out from 1st July to 30th September, 2012.

**Dietary treatments**

The laying hens were fed a basal diet supplemented with two levels of each Selenium (Sel- Plex) and chromium picolinate. Birds were assigned to each of the following diet treatments:

1 - Basal diet (without any additives and served as control).
2 - Basal diet + 0.3 mg Se /kg diet.
3 - Basal diet + 0.5 mg Se /kg diet.
4 - Basal diet + 1200 μg Cr /kg diet.
5 - Basal diet + 1400 μg Cr /kg diet.
6 - Basal diet + 0.3 mg Se + 1200 μg Cr /kg diet.
7 - Basal diet + 0.5 mg Se + 1200 μg Cr /kg diet.
8 - Basal diet + 0.3 mg Se + 1400 μg Cr /kg diet.
9 - Basal diet + 0.5 mg Se + 1400 μg Cr /kg diet

**Ambient temperature and relative humidity**

Daily ambient temperature (AT) and relative humidity (RH) were recorded inside the laying houses three times per day (at 8 a.m., 12 p.m. and 4 p.m.) and twice at night (at 12 a.m. and 4 a.m.) during the experimental period. Also, means of maximum and minimum AT and RH were recorded monthly (Table 2). Indoor climatic conditions were recorded using electronic digital thermo-hygrometer. Temperature-humidity index (THI) was calculated according to Marai et al. (2001). They established several stages of thermal comfort values such as: absence of heat stress (<27.8), moderate heat stress (27.8-28.8), severe heat stress (28.9-29.9) and very severe heat stress (>30.0), based on variations in the bird’s body temperature, using the following mathematical model:

\[
\text{THI} = \text{db}^\circ\text{C} - [(0.31 - 0.31 \times \text{RH}) \times (\text{db}^\circ\text{C} - 14.4)]
\]

Where: 
- THI = Temperature-humidity index.
- db°C = dry bulb temperature in centigrades.
- RH = relative humidity %.

**Temperature-humidity index**

During the experimental period (Summer 2012), birds exposed to very severe heat stress as the estimated THI values were 32.97, 33.39 and 31.06 during July, August and September, respectively (Table 2). In this study, the ambient temperature was higher than the recommended normothermia zone of 18-28°C (Donkoh, 1989; Holik, 2009) established for poultry in the tropical regions. This is a clear indication that the experimental layers, used in the present study, were subjected to a severe heat stress.

**Blood constituents of laying hens**

At 36 weeks of age, three hens per treatment were slaughtered within one to two hours post-oviposition in order to take some measurements. Blood samples of individual hens were collected from the jugular veins during slaughter into two heparinized tubes. The blood sample in the first tube was used for the measurement of some hematological characteristics. Improved Neubauer hemocytometer (Brand, Wertheim, Germany) was used for counting RBCs and WBCs according to Natt and Herrick (1952). The hemoglobin (Hb) content was determined using the Drabkin’s technique according to Dein (1984). Packed cell volume (PCV) was determined by using the micro hematocrit capillary tubes and the micro hematocrit centrifuge at 12,000 rpm for 5 min. according to Bara (1988). Blood films were stained by Giemsa stain and the differential leucocytic count was performed according to Jain (1986).

The second half of blood sample was centrifuged at 4000 r.p.m. for 15 minutes to separate blood plasma, then stored at −20 °C for determining the concentrations of plasma total protein (Gornall et al., 1949), albumin (Dumas
et al., 1971), creatinine (Bartles et al., 1972), urea (Fawcett and Socct, 1960), Cl (Schales and Schales, 1941), Ca++ (Moorehead and Biggs, 1974), inorganic P (Goldenberg and Fernandez, 1966), cholesterol (Richmond, 1973), glucose (Trinder, 1969), total lipids (Zollner and Kirsch, 1962), triglycerides (Fassati and Prencipe, 1982), and activities of plasma transaminases: aspartate aminotransferase (AST) and alanine aminotransferase (ALT) (Reitman and Frankel, 1957). Globulin concentration was obtained by subtracting the values of albumin from the corresponding value of total protein. At the same time, plasma concentrations of triiodothyronine (T3) and thyroxine (T4) were assayed according to methods described by Darras et al. (1992).

**Statistical analysis**

Data were subjected to statistical analysis by one-way analysis of variance using SPSS® statistical software (SPSS, 1999, version 10.0). Significant difference among treatment means were detected by Duncan’s Multiple Range Test (Duncan, 1955) at 5% level of significance at (P≤0.05). The following model was used to study the effect of Selenium (Sel-Plex) and chromium picolinate on blood parameters of laying hens as follows:

\[ Y_{ij} = M + T_i + e_{ij} \]

Where:
- \( Y_{ij} \) = observation for each dependent variable.
- \( M \) = overall mean.
- \( T_i \) = Treatment effects (i = 1, 2,...... and 9).
- \( e_{ij} \) = Random error.

**RESULTS AND DISCUSSION**

**Blood hematological variables**

Effects of dietary supplementation with Se and Cr and their combination on some blood hematological variables of 36-week-old laying hens reared under the summer hot climate are presented in (Table 3).

There were significant increases (P≤0.05) in total counts of blood leukocytes (WBCs), hemoglobin (Hb), packed cell volume (PCV) and total counts of blood erythrocytes (RBCs) as affected by dietary supplementations comparing to their control counterparts. When 0.5 mg Se plus 1200 μg/kg Cr was used total counts of blood leukocytes were comparable to that of the control. Also, treatments 1200 μg/kg Cr, 0.5 mg Se plus 1200 μg/kg Cr and 0.5 mg Se plus 1400 μg/kg Cr were similar with control in packed cell volume. Also, treatments 1200 μg/kg Cr and 0.5 mg Se separately or in combination with both levels of Cr in total counts of blood erythrocytes were similar with control value. However, there were significant decreases (P≤0.05) in monocytes cells compared to its counterpart in control group. Furthermore, lymphocytes cells were not affected by the addition of dietary supplementation, but lymphocytes were affected for hens given 1400 μg/kg Cr in their diet. Also, no significant differences were detected in heterophil cell due to the dietary supplementation.

It is well known that heat stress is one of the major stressors on poultry production which produces a wide range of physiological changes.
inhibiting effect of heat stress on blood concentrations of RBCs and Hb (Donkoh, 1989) and WBCs (Lin et al., 2003; Mashaly et al., 2004) is well documented in both broiler chicks and laying hens. Kutlu and Forbes (1993) found elevations in plasma concentrations of protein, glucose, heterophil, lymphocyte and heterophil/lymphocyte ratio and decreases in potassium ion concentration in response to thermal stress. On other hand Hanafy et al. (2009) reported that organic selenium significantly (P≤0.05) increased blood Hb, RBC’s and WBC’s. Dietary selenium and Cr and their different combination inhibit leukocyte migration in broilers (Swain and Johri, 2000).

**Blood biochemical variables**

The effects of inclusion of Se and Cr and their combination on some blood biochemical variables of 36-week-old laying hens reared under the summer hot climate are presented in Tables 4a and 4b. In general, the results of dietary supplementation with Se and Cr and their combinations in (Table 4), exhibited no significant differences (P≤0.05) in plasma concentrations of total protein, albumin, globulin, albumin: globulin ratio, creatinine, urea and chloride in response to dietary supplementation. But when the diets were supplemented with 0.3 mg Se plus 1200 μg/kg Cr or 0.3 mg Se plus 1400 μg/kg Cr plasma levels of total protein and globulin were significantly higher (P≤0.05) than those of the control group. However, albumin: globulin ratio was significantly lower (P≤0.05) for the hens fed the diet supplemented with 0.3 mg Se plus 1400 μg/kg Cr as compared to the control birds. Similarly, blood plasma urea levels were significantly decreased (P≤0.05) when hens fed diets supplemented with 1200 μg/kg Cr, 0.3 mg Se plus 1200 μg/kg Cr and 0.3 mg Se plus 1400 μg/kg Cr compared with their control counterparts. Feeding Cr-supplemented diets at levels of 1200 or 1400 μg/kg to laying hens under hot environment conditions led to significantly higher (P≤0.05) levels of plasma chloride compared with that of the control group. Finally, no significant differences were detected in plasma levels of albumin and creatinine due to the dietary supplementation. Dietary treatments, applied in the present study, significantly improved (P≤0.05) plasma calcium but did not significantly affect phosphorus except the combination of 0.3 mg Se by 1400 μg/kg Cr which achieved the highest value of plasma calcium and also significantly increased phosphorus.

Our observation in this study is in agreement with that reported by Ozbey et al. (2004) and Seyrek et al. (2004) who reported that concentrations of blood serum glucose, triglyceride and cholesterol were significantly higher whereas, levels of total protein and albumin were significantly lower in laying Japanese quails exposed to heat stress. Also, Nalini et al. (2008) reported that at 42-45 °C, serum, growth hormone, creatinine, urea, glucose, cholesterol, triglycerides, AST and ALT increased significantly (P<0.05) from respective control mean values. The mechanism of this alteration has been reported to be through increased generation of free radicals at the cell level. Several authors have documented that free radical generation affects blood serum metabolites of plasma total protein, cholesterol and glucose which is manifested in bird’s adaptation response through decreased production performance (Lin et al. 2005; Imik et al. 2009). In contrary, Koelkebeck and
Odom (1995) found that acute heat stress had no effect on blood plasma levels of glucose, total protein and creatinine in laying hens.

Organic selenium had more pronounced effect on total protein. Kim and Mahan (2003) indicated that selenium is biochemically similar to sulphur, selenium replaces the sulphur molecule in the normal biosynthetic pathways of the yeast cell and is absorbed actively across the intestine by the same amino acid carrier. Combs and Combs (1986) reported that supplemented organic Se to broiler breeders and layers was actively absorbed and can be directly incorporated into protein. In harmony with our results, Attia et al. (2010) found that vitamin E and/or Se supplementation significantly decreased triglycerides, albumin, globulin and albumin / globulin ratio on layer, however Hanafy et al. (2009) found that organic selenium increase albumin. The increase in plasma total protein concentration by Cr supplementation is in agreement with previous results obtained for laying Japanese quails (Sahin et al., 2001; Abdel-Mageed and Hassan, 2012). The positive effect of Cr on plasma protein and its fractions may be due to the anabolic action of insulin mediated through increasing the amino acids synthesis by liver, enhancement the incorporation of several amino acids into protein (Uyanik et al., 2002).

Results presented in Table 4b exhibited that laying hens fed diets supplemented with Se and Cr and their combinations had significantly lower (Ps<0.05 plasma levels of cholesterol, glucose, total lipids and triglycerides as well as activity of ALT and AST while plasma concentrations of triiodothyronine (T_3), and thyroxine (T_4) were significantly higher (Ps<0.05) compared with their control counterparts.

In agreement with the present results, Kalaba (2007) and Ibrahiem (2008) reported that stressful factors can increase serum levels of cholesterol, glucose, total lipids and triglycerides, and activity of transaminases (ALT and AST), but may decrease the plasma levels of the hormones: T_3 and T_4. According to Bhatti and Dil (2005), alteration in serum enzymes activity under stress conditions occur due to malfunctioning of liver, as degenerating and necrotic cells leak enzymes from cytoplasm.

According to the current results added dietary selenium alleviated the adverse effect of heat stress on the activity of AST and ALT in the blood plasma. In this direction, El-Mallah et al. (2011) observed a beneficial effect for selenium yeast on ALT and AST. The use of organic selenium (Sel-plex) as a source of supplemental dietary Se provides a highly efficient form of organic Se and facilitates a greater antioxidant enzymes which can readily reduce peroxides and other free radicals thereby compromising the cell membranes (Edens and Gowdy, 2005). On the other hand, Abaza (2002) observed that plasma cholesterol concentration was significantly increased by supplemental Se and/or vitamin E. However, Ljubic et al. (2006) suggested that the organic Se supplementation influences cholesterol metabolism in adipose tissue by decreasing the total cholesterol concentration during the fattening period and increasing the free cholesterol concentration after 48 h feed deprivation. Changes in enzymes responsible for regulating cholesterol synthesis, oxidation or elimination may be responsible for lowering the cholesterol synthesis in mature as well as
immature chickens (Konjufca et al., 1997). Also, Saito et al. (2007) demonstrated that the oxidative stress induced by selenium deficiency can enhance lipid and cholesterol peroxidation in cultured cells.

In agreement with the present findings, several studies have shown that dietary Cr supplementation decreased blood glucose and cholesterol concentrations in laying hens (Lien et al., 1996; Uyanik et al., 2002). Also, Abdallah et al. (2013) found that Cr picolinate administration significantly decreased serum cholesterol and glucose, and activity of ALT and AST. In addition, Ibrahim (2008) found the same results for laying hens reared under heat stress. According to Prasada and Gowda (2005) Cr act as glucose tolerance factor which can increase the uptake of glucose by cells and thus attenuating the insulin action. The reducing effect of Cr on plasma glucose in the present study may support this suggestion.

Table 1: The composition and chemical calculated analysis of the experimental diets.

<table>
<thead>
<tr>
<th>Ingredients</th>
<th>%</th>
</tr>
</thead>
<tbody>
<tr>
<td>Yellow corn</td>
<td>66.00</td>
</tr>
<tr>
<td>Soybean meal (44%)</td>
<td>24.00</td>
</tr>
<tr>
<td>Limestone</td>
<td>7.59</td>
</tr>
<tr>
<td>Di-calcium phosphate</td>
<td>1.71</td>
</tr>
<tr>
<td>Sodium chloride</td>
<td>0.30</td>
</tr>
<tr>
<td>Vit. &amp; Min. Mixture*</td>
<td>0.30</td>
</tr>
<tr>
<td>DL. Methionine</td>
<td>0.10</td>
</tr>
<tr>
<td>Total</td>
<td>100</td>
</tr>
<tr>
<td>Metabolizable energy (kcal/kg)</td>
<td></td>
</tr>
<tr>
<td>Crude Protein, %</td>
<td>16.43</td>
</tr>
<tr>
<td>Crude fiber, %</td>
<td>3.20</td>
</tr>
<tr>
<td>Ether extract, %</td>
<td>2.70</td>
</tr>
<tr>
<td>Calcium, %</td>
<td>3.33</td>
</tr>
<tr>
<td>Available phosphate, %</td>
<td>0.45</td>
</tr>
<tr>
<td>Total phosphate, %</td>
<td>0.66</td>
</tr>
<tr>
<td>Lysine, %</td>
<td>0.86</td>
</tr>
<tr>
<td>Methionine, %</td>
<td>0.39</td>
</tr>
</tbody>
</table>

*Supplied per kg of diet: vit.A, 10000 IU; D3, 2000 IU; Vit.E, 10mg; Vit.K3, 1mg; vit. B1, 1mg; vit. B2, 5mg; vit.B6, 1.5mg; vit. B12, 10mcg; Niacin, 30mg; Pantothenic acid, 10mg; Folic acid, 1mg; Biotin, 50µg; Choline, 260mg; Copper, 4mg; Iron; 30mg; Manganese, 60mg; Zinc, 60mg; Iodine, 1.3mg; Selenium, 0.1mg; Cobalt, 0.1mg.

Several researchers reported reduced blood plasma concentrations of T3 and T4 in heat-stressed chickens (Kalaba, 2007). The inverse relationship between plasma concentration of T3 and environmental temperature has been well known (Yahav, 1999). In this regard, Beckett et al. (1987) found that T3 is produced by 5′-deiodination of T4 particularly in the thyroid, liver and kidney. The activity of the selenoenzyme catalyzing 5′-deiodination (5′-ID) in rats is affected by Se deficiency (Kohrle et al., 1992). It has been reported that hepatic 5′-ID activity in the Se deficient rats is 10-fold lower but plasma
T₃ concentration is significantly lower in Se supplemented rats than in normal rats (Beckett et al., 1992) which is in accordance with the present results. Greater plasma concentrations of T3 and T4, reported in the present study, with higher dietary chromium could support a greater performance of laying hens, as T3 and T4 are considered as important growth promoters in animals (McNabb and King, 1993). Also, Sahin et al. (2001) reported similar results on Japanese quails.

From the present results, it is concluded that enhancing hen’s diets with selenium (as Sel-Plex) and chromium picolinate singly or in combination at levels of 0.3 mg Se plus 1400 μg Cr /kg diet can consider as an effective management practice for reducing the adverse effects of heat stress for laying hens.

Table (2): Means of indoor ambient temperature, relative humidity and temperature-Humidity Index measured within the open-sided laying house, during experimental period (from July 2012 to September 2012).

<table>
<thead>
<tr>
<th>Months</th>
<th>Ambient temperature (°C)</th>
<th>Relative humidity (%)</th>
<th>Temperature-Humidity Index</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>minimum</td>
<td>maximum</td>
<td>minimum</td>
</tr>
<tr>
<td>July</td>
<td>25.30 ±0.08</td>
<td>34.25 ±0.50</td>
<td>48.02 ±2.27</td>
</tr>
<tr>
<td>August</td>
<td>25.02 ±0.47</td>
<td>34.66 ±0.14</td>
<td>47.13 ±0.19</td>
</tr>
<tr>
<td>September</td>
<td>22.73 ±0.59</td>
<td>32.28 ±0.70</td>
<td>47.30 ±1.47</td>
</tr>
</tbody>
</table>

Table 3: Blood hematological variables of laying hens fed diets supplemented with Se, Cr or their combinations at 36 wks old.

<table>
<thead>
<tr>
<th>Items</th>
<th>WBCs 10³/mm³</th>
<th>Lymph %</th>
<th>Heter %</th>
<th>Mono %</th>
<th>PCV %</th>
<th>Hb gm/dl</th>
<th>RBCs 10⁶/mm³</th>
</tr>
</thead>
<tbody>
<tr>
<td>control</td>
<td>25.17 ±0.20</td>
<td>58.34 ±2.09</td>
<td>27.82 ±1.79</td>
<td>13.85 ±0.31</td>
<td>26.33 ±1.20</td>
<td>8.17 ±0.12</td>
<td>2.03 ±0.19</td>
</tr>
<tr>
<td>0.3 mg Se</td>
<td>28.57 ±0.82</td>
<td>64.33 ±5.36</td>
<td>25.52 ±4.75</td>
<td>10.14 ±0.94</td>
<td>30.67 ±2.19</td>
<td>10.47 ±0.43</td>
<td>2.67 ±0.29</td>
</tr>
<tr>
<td>0.5 mg Se</td>
<td>30.30 ±0.72</td>
<td>69.26 ±4.68</td>
<td>30.74 ±5.48</td>
<td>10.00 ±1.15</td>
<td>30.67 ±1.20</td>
<td>10.00 ±0.60</td>
<td>2.47 ±0.02</td>
</tr>
<tr>
<td>1200 μg/kg Cr</td>
<td>28.10 ±0.36</td>
<td>62.53 ±4.22</td>
<td>26.45 ±4.26</td>
<td>11.02 ±0.58</td>
<td>29.00 ±0.18</td>
<td>9.67 ±0.57</td>
<td>2.57 ±0.12</td>
</tr>
<tr>
<td>1400 μg/kg Cr</td>
<td>30.03 ±0.33</td>
<td>65.40 ±6.98</td>
<td>24.10 ±6.28</td>
<td>10.50 ±2.20</td>
<td>32.33 ±1.45</td>
<td>10.33 ±0.17</td>
<td>2.80 ±0.06</td>
</tr>
<tr>
<td>0.3 mg Se + 1200 μg/kg Cr</td>
<td>35.10 ±0.49</td>
<td>69.59 ±2.38</td>
<td>20.55 ±2.29</td>
<td>9.86 ±0.20</td>
<td>31.00 ±1.00</td>
<td>9.77 ±0.10</td>
<td>2.70 ±0.26</td>
</tr>
<tr>
<td>0.5 mg Se + 1200 μg/kg Cr</td>
<td>26.43 ±0.30</td>
<td>69.56 ±7.82</td>
<td>29.88 ±8.11</td>
<td>10.53 ±0.30</td>
<td>30.00 ±1.15</td>
<td>10.00 ±0.51</td>
<td>2.40 ±0.25</td>
</tr>
<tr>
<td>0.3 mg Se + 1400 μg/kg Cr</td>
<td>35.43 ±0.38</td>
<td>72.47 ±0.76</td>
<td>20.81 ±0.55</td>
<td>6.72 ±0.24</td>
<td>30.33 ±0.88</td>
<td>10.95 ±0.26</td>
<td>2.90 ±0.06</td>
</tr>
<tr>
<td>0.5 mg Se + 1400 μg/kg Cr</td>
<td>28.20 ±0.61</td>
<td>71.12 ±1.14</td>
<td>20.51 ±1.40</td>
<td>8.37 ±0.26</td>
<td>30.00 ±0.58</td>
<td>10.03 ±0.58</td>
<td>2.30 ±0.15</td>
</tr>
</tbody>
</table>

Significant * * * NS * * * *

* * means in the same column having different superscripts differ significantly at (P≤ 0.05).
NS (Not significant), * (Significant at P≤ 0.05).
Table (4a): Blood biochemical of laying hens fed diets supplemented with organic forms of Se, Cr or their combinations at 36 wks old.

<table>
<thead>
<tr>
<th>Items</th>
<th>Protein g/dl</th>
<th>Albumin g/dl</th>
<th>Globulin g/dl</th>
<th>A / G ratio</th>
<th>Creatine mg/dl</th>
<th>Urea mg/dl</th>
<th>Chloride mg/dl</th>
<th>Ca mg/dl</th>
<th>P mg/dl</th>
</tr>
</thead>
<tbody>
<tr>
<td>control</td>
<td>4.93±0.05</td>
<td>2.43±0.04</td>
<td>2.50±0.08</td>
<td>0.98±0.04</td>
<td>1.35±0.27</td>
<td>5.57±0.02</td>
<td>0.08±0.01</td>
<td>18.52±0.39</td>
<td>3.02±0.29</td>
</tr>
<tr>
<td>0.3 mg Se</td>
<td>5.85±0.37</td>
<td>2.35±0.26</td>
<td>3.50±0.43</td>
<td>0.70±0.14</td>
<td>0.79±0.28</td>
<td>4.12±0.06</td>
<td>0.08±0.01</td>
<td>27.64±0.51</td>
<td>3.83±0.67</td>
</tr>
<tr>
<td>0.5 mg Se</td>
<td>4.94±0.49</td>
<td>2.29±0.06</td>
<td>2.65±0.54</td>
<td>0.97±0.26</td>
<td>0.94±0.27</td>
<td>4.23±0.08</td>
<td>0.08±0.01</td>
<td>25.54±0.48</td>
<td>3.77±0.48</td>
</tr>
<tr>
<td>1200 μg Cr</td>
<td>5.13±0.37</td>
<td>2.18±0.11</td>
<td>2.95±0.48</td>
<td>0.80±0.17</td>
<td>1.24±0.21</td>
<td>3.94±0.11</td>
<td>0.11±0.01</td>
<td>25.55±0.40</td>
<td>3.04±0.02</td>
</tr>
<tr>
<td>1400 μg Cr</td>
<td>5.67±0.35</td>
<td>2.40±0.20</td>
<td>3.26±0.52</td>
<td>0.79±0.17</td>
<td>1.17±0.32</td>
<td>4.13±0.11</td>
<td>0.11±0.01</td>
<td>27.88±0.06</td>
<td>4.13±0.06</td>
</tr>
<tr>
<td>0.3 mg Se + 1200 μg Cr</td>
<td>6.08±0.33</td>
<td>2.54±0.11</td>
<td>3.54±0.34</td>
<td>0.73±0.08</td>
<td>0.77±0.21</td>
<td>3.79±0.08</td>
<td>0.08±0.01</td>
<td>29.49±0.38</td>
<td>4.46±0.23</td>
</tr>
<tr>
<td>0.5 mg Se + 1200 μg Cr</td>
<td>5.31±0.36</td>
<td>2.46±0.22</td>
<td>2.87±0.20</td>
<td>0.85±0.08</td>
<td>1.31±0.25</td>
<td>4.54±0.08</td>
<td>0.08±0.01</td>
<td>25.40±0.35</td>
<td>3.99±0.20</td>
</tr>
<tr>
<td>0.3 mg Se + 1400 μg Cr</td>
<td>6.29±0.38</td>
<td>2.27±0.04</td>
<td>3.99±0.34</td>
<td>0.58±0.04</td>
<td>1.18±0.11</td>
<td>3.63±0.09</td>
<td>0.09±0.01</td>
<td>30.51±0.32</td>
<td>4.73±0.32</td>
</tr>
<tr>
<td>0.5 mg Se + 1400 μg Cr</td>
<td>5.02±0.30</td>
<td>2.42±0.15</td>
<td>2.60±0.17</td>
<td>0.93±0.03</td>
<td>1.24±0.15</td>
<td>4.14±0.08</td>
<td>0.08±0.01</td>
<td>26.17±0.16</td>
<td>3.53±0.26</td>
</tr>
</tbody>
</table>

Significant: * NS a,b,c * * * * *

NS (Not significant), * (Significant at P≤ 0.05).

Table (4b): Blood biochemical of laying hens fed diets supplemented with organic forms of Se, Cr or their combinations at 36 wks old.

<table>
<thead>
<tr>
<th>Items</th>
<th>cholesterol mg/dl</th>
<th>glucose mg/dl</th>
<th>total lipids mg/dl</th>
<th>triglycerides mg/dl</th>
<th>ALT u/ml</th>
<th>AST u/ml</th>
<th>T₃ nmol/l</th>
<th>T₄ nmol/l</th>
</tr>
</thead>
<tbody>
<tr>
<td>control</td>
<td>203.43±1.26</td>
<td>242.37±0.24</td>
<td>22.07±1.87</td>
<td>391.64±34.27</td>
<td>52.67±5.33</td>
<td>65.33±6.07</td>
<td>1.18±0.10</td>
<td>12.70±1.12</td>
</tr>
<tr>
<td>0.3 mg Se</td>
<td>189.11±2.88</td>
<td>229.67±0.42</td>
<td>17.59±0.82</td>
<td>232.81±11.96</td>
<td>27.00±8.14</td>
<td>41.67±4.10</td>
<td>1.62±0.08</td>
<td>20.04±0.95</td>
</tr>
<tr>
<td>0.5 mg Se</td>
<td>174.58±0.76</td>
<td>230.30±0.52</td>
<td>17.13±0.52</td>
<td>238.89±17.94</td>
<td>30.67±5.67</td>
<td>37.67±3.28</td>
<td>1.32±0.03</td>
<td>17.90±1.19</td>
</tr>
<tr>
<td>1200 μg Cr</td>
<td>176.02±7.23</td>
<td>229.23±0.32</td>
<td>18.48±0.82</td>
<td>322.86±10.37</td>
<td>25.00±4.00</td>
<td>38.67±5.21</td>
<td>1.52±0.04</td>
<td>16.83±0.42</td>
</tr>
<tr>
<td>1400 μg Cr</td>
<td>158.69±2.31</td>
<td>224.17±3.26</td>
<td>17.39±1.13</td>
<td>216.43±17.38</td>
<td>25.00±4.33</td>
<td>43.33±1.76</td>
<td>1.53±0.08</td>
<td>18.89±0.58</td>
</tr>
<tr>
<td>0.3 mg Se + 1200 μg Cr</td>
<td>163.19±3.09</td>
<td>219.50±0.35</td>
<td>16.48±2.01</td>
<td>191.17±20.42</td>
<td>27.0±8.14</td>
<td>37.33±3.18</td>
<td>1.68±0.03</td>
<td>20.72±0.25</td>
</tr>
<tr>
<td>0.5 mg Se + 1200 μg Cr</td>
<td>177.14±9.86</td>
<td>228.30±15.48</td>
<td>15.48±6.21</td>
<td>221.34±26.57</td>
<td>36.33±5.67</td>
<td>36.00±0.00</td>
<td>1.32±1.75</td>
<td>15.55±0.58</td>
</tr>
<tr>
<td>0.3 mg Se + 1400 μg Cr</td>
<td>152.42±2.79</td>
<td>218.37±0.41</td>
<td>13.17±0.03</td>
<td>212.92±21.72</td>
<td>21.33±3.87</td>
<td>27.67±3.18</td>
<td>1.75±0.10</td>
<td>20.96±0.96</td>
</tr>
<tr>
<td>0.5 mg Se + 1400 μg Cr</td>
<td>184.49±2.33</td>
<td>226.97±1.72</td>
<td>14.20±1.61</td>
<td>254.09±5.67</td>
<td>40.00±3.46</td>
<td>40.00±1.58</td>
<td>1.58±0.00</td>
<td>23.00±0.51</td>
</tr>
</tbody>
</table>

Significant: * NS a,b,c * * * * *

NS (Not significant), * (Significant at P≤ 0.05).

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REFERENCES


تخفيض التأثيرات العكسية للإجهاد الحراري على الدجاج البياض المحلى باستخدام السلينيوم والكروم العضويين.

المراجعات

السيرة حسن علي أبو عجلة زيد محمد العوضي قلبة أحمد عباس حسين طلبة و

ماجد عبد النبي أسماعيل أبيض

قسم أنثاق الدواجن-كلية الزراعة-جامعة المنورة.

قسم تربية الدواجن-معهد البحوث الحيواني-مركز البحوث الزراعية-الدقهلية.

أجرت هذه الدراسة في محطة بحوث الإنتاج الحيواني بخان كفر الشيخ خلال موسم صيف 2012 (يوليو-أغسطس-سبتمبر) ليبحث تأثير إضافة السلينيوم والكروم العضويين على تأثير إجهاد الحرارة على بعض خصائص الدم أثناء ارتفاع درجات الحرارة خلال فصل الصيف في مصر. استخدم في هذه الدراسة 271 دجاجة من سلالة الناصية المستنبطة محليا عند عمر 24 أسبوع وقد وزعت الطيور عشوائيا في أバランス جماعية إلى تسع مجموعات (كل مجموعة 30 دجاجة) كانت تتكون من ثلاث مكررات (كل مكررة 10 دجاجات) وقد تكون الغذاء التجريب في إضافة الغذاء الأساسي من سلطة تكوينية من كل من السلينيوم سلبلكس (1.0 ملليجرام/كجم) وبيكولينات الكروم (0.0211 جزء في المليون/كجم). واستمرت التجربة من عمر 22 إلى 32 أسبوع وكانت أهم النتائج كالآتي:

- أدت الاضافات لزيادة معنوية في عدد كرات الدم البيضاء بانواعها وكذلك الهيموجلوبين وحجم الخلايا وعدد كرات الدم الحمراء مقارنة بالمعاملة الكنترول.
- كان هناك تحسن أيضاً في محتوى الدم من البروتين الكلي والليبيدات والجلوبيولين والكرياتين والكالسيوم، حيث كانت أفضل النتائج عند إضافة 1.0 ملليجرام سلينيوم مع 0.0211 جزء في المليون كروم لل العليقة.
- كان هناك نقص في محتوى الدم من ALT وAST نتيجة إضافة كروم وسلينيوم إلى العليقة، حيث كانت أفضل النتائج عند إضافة 1.0 ملليجرام سلينيوم مع 0.0211 جزء في المليون كروم لل العليقة، ونتيجة إضافة 1.0 ملليجرام سلينيوم مع 0.0211 جزء في المليون كروم لل العليقة.

يعتبر من النتائج أن إضافة بيكولينات الكروم والسلنيوم سلبلكس خاصة بعد 3.0 ملليجرام سلينيوم مع 1400 جزء في المليون كروم يمكن أن يستخدم في مقاومة التأثيرات السلبية للحرارة العالية خلال فصل الصيف من خلال تحسين بعض خصائص الدم للدجاج البياض.

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