INTERLEUKIN-1 RECEPTOR ANTAGONIST GENE POLYMORPHISM AND OXIDATIVE STRESS IN CHRONIC HEPATITIS C VIRUS INFECTION.

¹Abdel-Aziz, A.F. : ² El-Said, A.M. ; ³ Abd El-Hafiz, Shaimaa. and ⁴ Settin, A.A.

¹Department of chemistry, Biochemistry Division, Faculty of Science, Mansoura University, Egypt (<u>afaziz@mans.edu.eg</u>); ²Genetics Unit, Faculty of Medicine, Mansoura University, Egypt (<u>dr_afaf96@yahoo.com</u>); ³Specialist of biochemistry, Researcher in Children Hospital, Mansoura University. Egypt <u>dr.shosho2007@gmail.com</u>); ⁴Departement of Pediatrics, Faculty of Medicine, Mansoura University, Egypt (<u>settin60@gmail.com</u>).

Received : (8 /3 / 2011)

ABSTRACT

Cytokines like interleukin-1 receptor antagonist have an important role in the defense against viral infection. These cytokines are thought to play a central role in liver metabolism and in the immune response to viral agents. HCV infection is characterized by a systemic oxidative stress that is most likely caused by a combination of chronic inflammation, iron overload, liver damage, and proteins encoded by HCV. This work was planned in order to check the association of genetic polymorphism of interleukin-1 receptor antagonist (IL-1Ra) gene with chronic hepatitis C infection and its relation to oxidative stress markers.

The polymorphism analyzed, IL-1Ra genotype A1A2 showed lower significant frequencies in YICV patients when compared with controls (52.7% vs. 71.0%, p = 0.006). This means that the genotype A1A2 might be considered a protective genotype against HCV infection. On the other hand, the frequency of A2A2 genotypes showed a higher significance in HCV patients than controls (15.5% vs. 8.1%, p=0.02) and A1A1 genotype frequency was higher in patients when compared with controls (27.3% vs. 20.2%, p=0.26),

Regarding to oxidative stress markers, in HCV patients showed significantly lower mean level of total antioxidant capacity (TAC) (p < 0.001), while these patients have significantly higher level of malonyldialdehyde (MDA) (p < 0.001). On the other hand, comparing cases-subgroups in terms of their assigned IL-1Ra genotypes, nitric oxide (NO) mean level was significantly higher in patients having the A2A2 genotype than other patients. Finally, the present study suggested that, the patients having the IL-1Ra A2A2 genotype were at higher risk for getting HCV infection and liver complications whereas those with A1A2 genotype seemed to be more protected against such infection. In addition, MDA, TAC and NO' levels might be used as monitoring markers for oxidative stress in HCV cases with IL-1Ra A2A2 genotype.

Key words: Hepatitis C virus; chronic liver disease; IL-1 Receptor antagonist ; nitric oxide; total antioxidant capacity; malonyldialdehyde; Polymerase Chain Reaction.

Corresponding author: Abdel-Aziz, A.F., Department of Chemistry, Biochemistry Division, Faculty of Science, Mansoura University, Egypt (afaziz@mans.edu.eg;azizafaziz@hotmail.com)

INTRODUCTION

According to World Health Organization (WHO) data, 3% of human population, i.e. approximately 170 million people, is infected with HCV. Chronically infected individuals serve as a source of transmission to others and are at risk for severe HCV related diseases, such as liver cirrhosis and hepatocellular carcinoma, during the decades after initial infection [Brass et al., (2006)]. Therefore, it is important to estimate the demographic history of HCV infection, to predict the future burden of disease. Egypt is the highest prevalence of hepatitis C in the world and the national prevalence rate of HCV antibody positively has been estimated to be between 10-13% [Mohamed (2004)].

Cytokines are thought to have a central role in liver metabolism and in the immune response to viral agents [Gramantieri et al., (1999)]. These cytokines have an important role in defense against viral infection, indirectly through determination of the predominant pattern of the host response, and directly through inhibition of viral replication [Koziel (1999)]. In fact, serum levels of cytokines are elevated in patients with chronic hepatitis C, suggesting their role in inflammation of the liver [Falasca et al., (2006) and Gramenzi et al., (2005)].

The heritable differences in cytokine gene expression have been proposed as risk factors for the development of liver cirrhosis in chronic hepatitis C [Poynard et al., (2001)]. Genes encoding interlukin-1 (IL-1) are located on the 430 kb region of chromosome 2q13-21, consisting of three homologous genes: IL-1A, IL-1 β and interlukin-1 receptor antagonist (IL-1Ra) [Nemetz et al., (1999)]. The IL-1 Receptor antagonist (IL-1Ra) is one member of the IL-1 family that represents an agent which binds to the same receptor on the cell surface as IL-1, and thus prevents IL-1 from sending a signal to that cell [Bahr et al., (2003)].

In hepatitis C, more severe grades of inflammation and fibrosis are associated with a shift in the balance between the hepatic expression of interleukin IL-1 β and interlukin

108

1-receptor agonist (IL-1Ra) towards more IL-1 β activity [Gramantieri et al., (1999)] and genetic polymorphism for both IL-1 β and IL-1Ra may increase the risk of HCV patients to develop cirrhosis [Bahr et al., (2003)].

IL-1Ra contains an 86 bp variable number tandem repeat (VNTR) polymorphism in intron 2 [Garcia-Gonzalez et al., (2001)]. These polymorphisms are located within the regulatory regions of the genes and have potential functional importance by modulating IL-1 protein production, and are related to the development of some diseases [Moos et al., (2001)]. There were five alleles in humans, including Allele 1 (four repeats, 410 bp), allele 2 (two repeats, 240 bp), Allele 3 (five repeats, 500 bp), allele 4 (three repeats, 325 bp) and Allele 5 (six repeats, 595 bp) [Zhang et al., (2004)].

HCV infection is characterized by a systemic oxidative stress that is most likely caused by a combination of chronic inflammation, iron overload, liver damage, and proteins encoded by HCV. The increased generation of reactive oxygen and nitrogen species, together with the decreased antioxidant defense, promotes the development and progression of hepatic and extra hepatic complications of HCV infection [Choi et al., (2004)]. Progression of liver disease was found to be linked with factors including modification of the immune response [Eggers et al., (2006)] and activation of proinflammatory cytokines both by virus C infection [Castellano-Higuera et al., (2008)].

Oxidative stress occurs due to discordance in balance between pro-oxidants and antioxidants. Normally, a rise in oxidative stress concomitantly enhances the anti-oxidative activity to protect the cell from damage. Chronic exposure to increased levels of oxidative stress may results in excess of reactive oxygen species within the hepatocytes [Pessayre et al, (2001)]. Oxidative stress can be described as a condition resulting from an uncontrolled increase in free oxygen radicals or an insufficiency in the antioxidant system under certain pathological states [Sariçam et al., (2005)].

Hepatitis C leads to up-regulation of hepatic inducible nitric oxide synthase (iNOS) gene expression and given that liver inflammation with hepatocellular damage, and fibrosis, it is likely that NO could be an active mediator inducing liver injury and carcinogenesis [Majano et al., (2003)]. Liver tissue from HCV-infected patients was shown to express elevated levels of iNOS transcripts compared with non-HCV-infected patients [Pârvu et al., (2005)].

SUBJECTS AND METHODS

This study was performed on 110 patients with hepatitis C virus (HCV). They were recruited from the Department of Tropical Medicine, Mansoura University Hospital, Egypt during the time of January 2009 to February 2010. These patients were 80 males and 30 females aged from 26.0 to 60.0 years. Their mean age were 43.3 ± 6.4 years. They were all positive for HCV antibody and HCV RNA that was detected quantitively by real time PCR (RT- PCR). These cases include 43.0 (31.9%) of them gave a history of Bilharziasis while 26 (19.3%) were complicated with liver cirrhosis manifested with ascietis. The control subjects including 124 healthy blood donors were taken as controls. They did not suffer from any liver disorders or other infectious.

allergic or autoimmune diseases. They were in the form of 110 males and 14 females with age ranging between 17.0 - 42.0 years with a mean \pm SD 26.0 \pm 5.7 years and a median age of 25.0 years (Table 1). An authorized approval was obtained from Mansoura University Scientific and Ethical Committees, in addition to an informed consent that was taken from all participants before the study.

Parameters	HCV patients (n=110)	Controls (n=124)
Age (years)	26.0 to 60.0	17.0 to 42.0
Mean ± SD	43.3 ± 6.4	26.0 ± 5.7
Median	44.0	25.0
Sex (M/F)	80/30	110/14
Ascietis	26.0 (19.3%)	0.0 (0.0%)
Bilharziasis	43.0 (31.9%)	0.0 (0.0%)
Smoking	24.0 (17.8%)	0.0 (0.0%)

Table (1): Demographic data of all studied HCV patients and controls.

DNA extraction and amplification of IL-1Ra intron 2:

For all cases and controls, DNA was extracted and purified using the generation DNA purification capture column kits (Gentra system, USA). Amplification via PCR technique was carried out for IL-1Ra intron 2 contained the 86 bp VNTR. Each PCR was carried out in 25 μ L reaction mixture containing 10 μ l Master Mix (Fermentas, Germany), 8 μ l PCR grade water, 2 μ l IL-1fa primer, 2 μ l IL-1ra primer (Bio Basic Inc., Canada) and 3 μ l extracted DNA. Primer sequence was (F): 5'-TCCTGGTCTGCAGGTAA-3', (R): 5'-CTCAGCAACACTCCTAT-3' [Kanemoto et al.,(2000)].

PCR conditions were as follows: initial denaturation cycle of 95°C for 5 min followed by 35 cycles in the form of 94°C for 30 s (denaturation), 55°C for 30 s (annealing) and 72°C for 1 min (extension) with a final extension cycle of 5 min at 72°C. Amplified segment was analyzed using Agarose gel (3%) electrophoresis and photographed on ultraviolet transilluminator [Zhang et al., (2004)].

Determination of biochemical markers related to Oxidative Stress:

Colorimetric determination of nitric oxide (NO') was done using nitrite assay kits (Biodiagnostic, Cairo, Egypt) through the method described by [Montgomery et al., (1961)] and determination of total antioxidant capacity (TAC) was done by using a commercially available kit (Biodiagnostic, Cairo, Egypt) through the method described by [Koracevic et al., (2001)], while estimation of serum malonyldialdehyde (MDA) was carried out through thiobarbituric acid (TBA) test using the method described by [Draper and Hadley, (1990)].

STATISTICAL ANALYSIS

Statistical analysis was done using the statistical package of social science (SPSS) software version 13. Gene frequency was determined by the gene counting method. Comparison between groups was done using Chi square test (χ 2) in addition to Odds ratio and 95% confidence intervals for the frequencies of studied alleles and genotypes; and student t-test for the numeric variables as oxidative stress markers. Probability (*p*) value was considered significant at a level ≤ 0.05 [SPSS, 1999].

RESULTS

In the present study, 4 alleles and 5 genotypes of IL-1Ra gene could be recognized among Egyptian cases with HCV infection and normal controls, including Allele 1 (four repeats, 410 bp), allele 2 (two repeats, 240 bp), Allele 3 (five repeats, 500 bp), allele 4 (three repeats, 325 bp) and Allele 5 (six repeats, 595 bp) (Figure 1).

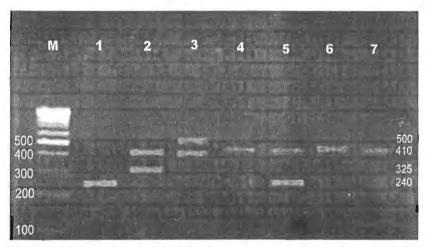


Fig (1): PCR product for IL-1R^a intron 2 gene polymorphism in HCV patients.

Lane (M): show DNA molecular mass marker 100 bp.

Lane (1): showed homozygote polymorphism of IL-1Ra A2/A2 genotype where A2 at 240 bp.

Lane (2): showed heterozygote polymorphism of IL-1Ra A1/A4 genotype where A1 at 410 bp and A4 at 325 bp.

Lane (3): showed heterozygote polymorphism of IL-1Ra A1/A3 genotype where A1 at 410 bp and A3 at 500 bp

Lane (4,6,7): showed homozygote polymorphism of IL-1Ra A1/A1 genotype where A1 at 410 bp.

Lane (5): showed heterozygote polymorphism of IL-1Ra A1/A2 1cy genotype where A1 at 410 bp and A2 at 240 bp.

ofA

that the genotype A1A2 might be considered a protective genotype against HCV infection.

On the other hand, the frequency of A1A1 and A2A2 homozygote genotype was higher in HCV patients than control group but is statistically insignificant (27.3% vs. 20.2% and 15.5% vs. 8.1% respectively, p>0.02) Table (2).

	Groups				
IL-1Ra	Ferrer	Control n=124 (%)	χ2 (p)	OR (95% CI)	
Genotypes					
AIAI	30 (27.3%)	25 (20.2%)	1.3 (0.26)	1.5 (0.81-2.7)	
A1A2	58 (52.7%)	88 (71.0%)	7.5 (0.006)*	0.4 (0.27-0.78)	
A2A2	17 (15.5%)	10 (8.1%)	2.4 (0.02)*	2.1 (0.91-4.77)	
A1A3	4 (3.6%)	1 (0.7%)	-	-	
A1A4	1 (0.9%)	0 (0.0)%	-		
Alleles	n=220 (%)	n=248 (%)			
A1	123 (55.9)	139 (56.0%)	0.01 (0.95)	0.99 (0.7-1.43)	
A2	92 (41.8%)	108 (44.0%)	0.08 (0.78)	0.93 (0.65-1.35)	
A3	4 (1.8%)	1 (0.34%)	-	-	
A4	1 (0.4%)	0 (0.0%)	-	-	

Table (2): Study of interleukin-1 receptor antagonist (IL-1Ra) in patients with HCV and controls.

IL-IRa genotypes are AIA1, AIA2, A2A2, AIA3 and AIA4. OR= odds ratio, 95% CI= 95% confidences interval,

 χ^2 = Chi square test.* p significant <0.05, (-) test is not applicable.

Patients with HCV infection associated with ascietis and liver cirrhosis showed a higher frequency of A2A2 genotype with a high odds ratio of 3.4 (95% CI = 1.2-10.4) compared with the cases having HCV without cirrhosis or ascietis, while patients with a history of bilharziasis showed the same results of significantly lower frequency of A1A2 genotype and non-significantly higher frequency of the other genotypes A1A1 and A2A2 compared to controls Table (3).

	AIAI	A1A2	A2A2
Controls (n=124)	25 (20.2%)	88 (71.0%)	10 (8.1%)
Bilharziasis (n=43)	13 (37.1%)	13 (37.1%)	7 (20.0%)
χ2 (p)#	1.3 (0.3)	20.5 (<0.0001)**	1.5 (0.22)
OR (95% CI)	1.72 (0.78-3.8)	0.2 (0.1-0.4)	2.2 (0.8-6.3)
Ascitis (n=26)	5 (19.2%)	13 (50.0%)	6 (23.1%)
$\chi^2(p)$	0.02 (0.87)	3.3 (0.06)	3.6 (0.05)*
OR (95% CI)	0.94 (0.32-2.7)	0.41 (0.2-0.98)	3.4 (1.2-10.4)*

Table (3): IL-1Ra genotypes in patients of HCV with Bilharziasis and Ascietis compared to controls.

Chi square test comparing cases subgroups with bilharziasis and ascities to controls, *p significant at 0.05 level, **p significant at <0.001 level.

Table (4) showed a significant decrease of serum activities of total antioxidant capacity (TAC) of patients with HCV infections $(1.10 \pm 0.10 \text{ mM/L})$ compared to controls value $(1.35 \pm 0.46 \text{ mM/L})$ (p = <0.001). While these patients have highly significant value with malonyldialdehyde $(15.23 \pm 3.28 \text{ nmol/ml})$ compared to control group $(7.92 \pm 2.74 \text{ nmol/ml})$ (p = <0.001).

Table (4): Total antioxidant capacity, nitric oxide and malonyldialdehyde of all studied HCV cases and controls.

Parameters	HCV patients M ± SD	Controls M ± SD	P value <0.05
TAC (mM/L)	1.10 ± 0.15	1.35 ± 0.46	0.001**
MDA (nmol/ml)	15.23 ± 3.28	7.92 ± 2.74	0.001**
NO (µmol)	22.82 ± 9.26	20.56 ± 6.66	0.17

TAC=Total Antioxidant Capacity, NO=Nitric Oxide, MDA=Malonyldialdehyde. Significant *p<0.05, highly significant **p<0.001.

The mean level values of TAC was shown to affect cases carrying the IL-1Ra A1A2 genotype $(1.13 \pm 0.09 \text{ vs. } 1.63 \pm 0.59, p= 0.01)$ while cases carrying the A1A1 and A1A2 genotypes showed significantly higher levels of MDA than controls $(15.22 \pm 213 \text{ vs. } 8.46 \pm 2.49, p=0.001)$ and $(14.76 \pm 3.58 \text{ vs. } 8.72 \pm 2.31, p=0.001)$ respectively. The mean level of NO' among HCV patients assigned IL-1Ra genotypes were significally elevated in patients having the A2A2 genotype followed by those having A1A2 and A1A1 genotypes (p=0.036) Table (5).

Parameters and IL-1Ra	HCV patients	Control
genotypes	$M \pm SD$	$M \pm SD$
TAC (mM/L)		
AIAI	$1.13 \pm .08$	1.24 ± 0.56
A1A2*	1.13 ± 0.09	1.63 ± 0.59
A2A2	1.18 ± 0.01	1.32 ± 0.11
MDA (nmol/ml)		
A1A1**	15.22 ± 2.13	8.46 ± 2.49
A1A2**	14.76 ± 3.58	8.72 ± 2.31
A2A2	17.09 ± 2.33	7.83 ± 5.38
NO (µmol)		+
AIAI	15.48 ± 3.44	22.63 ± 10.56
A1A2	22.23 ± 10.85	21.37 ± 5.85
A2A2#	29.64 ± 8.15	28.07 ± 7.55

Table (5): Serum activities of TAC (mM/L), NO (µmol), and MDA (nmol/ml) of HCV infected patients and control groups regarding IL-1Ra genotypes.

IL-IRa genotypes are AIAI, AIA2 and A2A2. TAC=total antioxidant capacity, NO=nitric oxide, MDA=malonyldialdehyde. # NO concentration was significantly higher among cases with IL-IRa A2A2 than AIA1 type, *p significant <0.05, **p significant <0.001.

DISCUSSION

Cytokines play an important role in liver metabolism and in the immune response to viral agents [Akpolat *et al.*, (2005)]. In this study, 4 alleles and 5 genotypes of IL-IRa gene could be recognized among Egyptian cases with HCV infection and normal controls. This genotype, A2A2 was the least frequent among Egyptian control subjects while the most frequent genotype was the heterozygosity genotype A1A2 that was significantly lower in patients with HCV compared to controls. On the other hand, homozygosity genotypes particularly of A2A2 was found higher among cases of HCV than controls mounting to a significant level among cases with ascietis due to liver cirrhosis. The same distribution was also found among cases of HCV associated with

bilharziasis. This may lead to speculate that homozygosity for the rare allele A2 is probably a risk genotype not only for liver affection with HCV and bilharziasis but also for the progression towards cirrhosis and ascietis. In contrast, heterozygosity genotype A1A2 seems to be probably protective against such affection.

Similarly, A2 allele was observed in about 10 to 30% of the Caucasian control population [Bahr et al., (2003)]. Also, [Wang et al., (2003)] found that the IL-1Ra A2 allele was less common in Japanese population (heterozygotes 5% to 9%; homozygotes, 0% to 2%). Regarding the homozygous genotype A1A1, it was found to be higher among Egyptian HCV cases (27.3%) compared to controls (20.2%), while [Constantini et al., (2002)] found that IL-1RN A1/A1 genotype was found in 66% of Northern European Caucasoid adult HCV patients but only 45% in HCV patient from Poland. Also, [Gramantieri et al., (1999)] found that among Italian population the IL-1Ra A2 allele was significantly more prevalent in patients with HCV-induced cirrhosis as compared to patients with chronic hepatitis C virus and controls. These points indicted that the role of IL-1Ra in the progression from chronic hepatitis to cirrhosis. In contrast, [Wang et al., (2003) and Tanaka et al., (2003)] stated that the IL-1RN A2A2 genotype had no association with liver disease progression, perhaps because it was less common in Japanese individuals (heterozygote 5% to 9%; homozygote, 0% to 2%).

In the present study we found that, no significant differences observed with respect to the rare IL-1Ra allele A3 and IL-1Ra allele A4 in Egyptian cases and controls. This observation was in agreement with data reported by [Glas et al., (2005)] in German population and by [Zhang et al., (2004)] in Chinese.

HCV infection was reported to be associated by increased markers of oxidative stress. [Choi et al., (2004)] found that lipid peroxidation products were increased in serum and liver specimen from hepatitis C patients. Nitric oxide (NO') is overproduced in liver cirrhosis and its disturbances seem to play a key role in the pathogenesis of chronic liver disease and are proposed as one of the major endogenous vasodilators in portal hypertension [Xu et al.,(2008)]. Under conditions of oxidative stress, as seen in certain chronic inflammatory disorders '.icluding hepatitis C, reactive NO' Species (RNOS), such as peroxynitrite and nitrogen oxides, are currently considered as the main mediators of the deleterious effects to the host of NO., including cytotoxicity and DNA damage [Zamora et al., (2000)]. It has been reported that patients with degenerative liver disease had increased lipid peroxide levels in liver tissue and serum [Seronello et aL, (2007)]. Exposure to more oxidant stress or an inability in the oxidative capacity of the cells might lead to acceleration of peroxidation reactions of some cellular molecules including lipids [Koruk et al., (2002)].

In this study patients infected with HCV showed a significant lower mean level of total antioxidant capacity (TAC) and a significant higher level of malonyldialdehyde (MDA), compared to controls. These observations are in agreement with [Vendemiale et al., (2001) and Aksoy et al., (2003)].

Moreover, [Paradis et al., (1997)] found a direct relationship between fibrosis and liver MDA in patients with HCV infection. Hassan et al., (2002)] reported increased NO levels via increased inducible NO synthase (iNOS) among Egyptian cases. We found that no significant correlation between nitrate levels and age or sex distribution in our cases, these observations are in accordance with El-Sherif et al., (2008)] who did not find any significant difference between nitrate levels in patients with chronic active hepatitis and healthy controls among Egyptian subjects. This was also consistent with [Pârvu et al., (2005)] among Ramanian and with [Arkenau et al., (2002)] among German subjects.

In this study the mean total levels of TAC and MDA showed insignificant difference among the cases with the 3 main genotypes but only NO' level showed a higher significant difference in cases with A2A2 genotype followed by A1A2 and A1A1 genotypes. [Castellano-Higuera et al., (2008)] found that cytokine levels contributed to the progression of chronic hepatitis C infection in Spanish population, also oxidative stress and cytokine secretion are closely related to each other's. Inflammatory conditions, such as infection or any situation leading to an acute organic stress, trigger a cytokine response and enhance oxidative stress.

In conclusion, we speculated that subjects having the IL-1Ra A2A2 genotype were having higher risk for getting HCV infection and liver complications whereas those with A1A2 genotype seemed to be more protected against such infections. Also, oxidative stress was documented in HCV cases and was manifested with high levels of MDA and low levels of TAC. In addition, NO' levels were higher among HCV cases with IL-1Ra A2A2 genotype than other cases.

REFERENCE

Akpolat N, Yahsi S, Godekmerdan A, Demirbag K, Yalniz M (2005): Relationship between serum cytokine levels and histopathological changes of liver in patients with hepatitis B. World J Gastroenterol; 7;11(21):3260-3.

Aksoy H, Taysi S, Altinkaynak K, Bakan E, Bakan N, and Kumtepe Y (2003): Antioxidant potential and transferrin, ceruloplasmin, and lipid peroxidation levels in women with preeclampsia. J Investig Med; 51(5): 284-287.

Arkenau HT, Stichtenoth DO, Frölich JC, Manns MP, and Böker KH (2002): Elevated nitric oxide levels in patients with chronic liver disease and cirrhosis correlate with disease stage and parameters of hyperdynamic circulation. Z Gastroenterol; 40(11):907-913.

Bahr MJ, el Menuawy M, Boeker KH, Musholt PB, Manns MP, and Lichtinghagen R (2003): Cytokine gene polymorphisms and the susceptibility to liver cirrhosis in patients with chronic hepatitis C. Liver Int 23: 420-425.

Brass V, Moradpour D, Blum HE (2006): Molecular virology of hepatitis C virus (HCV): 2006 update. Int J Med Sci.3(2):29-34.

Castellano-Higuera A, González-Reimers E, Alemán-Valls MR, Abreu-González P, Santolaria-Fernández F, De La Vega-Prieto, MJ, Gómez-Sirvent, JL, and Pelazas-González R (2008): Cytokines and lipid peroxidation in alcoholics with chronic hepatitis C virus infection. Alcohol Alcohol; 43(2):137-142.

Choi J, Lee KJ, Zheng Y, Yamaga AK, Lai MM, and Ou JH (2004): Reactive oxygen species suppress hepatitis C virus RNA replication in human hepatoma cells. Hepatology; 39(1):81-89.

Constantini PK, Wawrzynowicz-Sycrowska M, Clare M, Boron-Kaczmarska A, McFarlane IG, Cramp ME, and Donaldson PT (2002): Interleukin-1, interleukin-10 and tumour necrosis factor-alpha gene polymorphisms in hepatitis C virus infection: an investigation of the relationships with spontaneous viral clearance and response to alpha-interferon therapy. Liver; 22(5):404-412.

Draper HH, and Hadley M (1990): Malondialdehyde determination as index of lipid peroxidation. Methods Enzymol;186: 421-431.

Eggers V, Pascher A, Althoff H, Thiele S, Mütze J, Selignow J, Neuhaus P, and Spies CD (2006): Immune reactivity is more suppressed in patients with alcoholic liver disease than in patients with virus-induced cirrhosis after CRH stimulation. Alcohol Clin Exp Res; 30(1):140-149.

El-Sherif AM, Abou-Shady MA, Al-Bahrawy AM, Bakr RM, and Hosny AM (2008): Nitric oxide levels in chronic liver disease patients with and without oesophageal varices. Hepatol Int; 2(3):341-345.

Falasca K, Ucciferri C, Dalessandro M, Zingariello P, Mancino P, Petrarca C, Pizzigallo E, Conti P, Vecchiet J (2006): Cytokine patterns correlate with liver damage in patients with chronic hepatitis B and C. Ann Clin Lab Sci. Spring;36(2):144-150.

Garcia-Gonzalez MA, Lanas A, Santolaria S, Crusius JB, Serrano MT, and Peña AS (2001): The polymorphic IL-1B and IL-1RN genes in the aetiopathogenesis of peptic ulcer. Clin Exp Immunol;125(3):368-375.

Glas J, Török HP, Schneider A, Brünnler G, Kopp R, Albert ED, Stolte M. and Folwaczny C (2005): Allele 2 of the interleukin-1 receptor antagonist gene is associated with early gastric cancer. Erratum in: J Clin Oncol; 23(1):248.

Gramantieri L, Casali A, Trerè D, Gaiani S, Piscaglia F, Chieco P, Cola B, and Bolondi L (1999): Imbalance of IL-1 beta and IL-1 receptor antagonist mRNA in liver tissue from hepatitis C virus (HCV)-related chronic hepatitis. Clin Exp Immunol; 115(3):515-520.

Gramenzi A, Andreone P, Loggi E, Foschi FG, Cursaro C, Margotti M, Biselli M, Bernardi M (2005): Cytokine profile of peripheral blood mononuclear cells from patients with different outcomes of hepatitis C virus infection. J Viral Hepat; 12(5):525-30.

Hassan MI, Kassim SK, Ali HS, Sayed el-DA, and Khalifa A (2002): Evaluation of nitric oxide (NO) levels in hepatitis C virus (HCV) infection: relationship to schistosomiasis and liver cirrhosis among Egyptian patients. Dis Markers; 18(3):137-142.

Kanemoto K, Kawasaki J, Miyamoto T, Obayashi H, and Nishimura M (2000): Interleukin (IL)1beta, IL-1alpha, and IL-1 receptor antagonist gene polymorphisms in patients with temporal lobe epilepsy. Ann Neurol; 47(5):571-574.

Koracevic D, Koracevic G, Djordjevic V, Andrejevic S, and Cosic V (2001): Method for the measurement of antioxidant activity in human fluids. J Clin Pathol; 54(5):356-361.

Koruk M, Aksoy H, Akçay F, Onuk MD (2002): Antioxidant capacity and nitric oxide in patients with hepatic cirrhosis. Ann Clin Lab Sci. 32(3):252-6.

Koziel MJ, (1999): Cytokines in viral hepatitis. Semin Liver Dis; 19(2):157-169.

Majano PL, and Garcia-Monzon C (2003): Does nitric oxide play a pathogenic role in hepatitis C virus infection? Cell Death Differ; 10 Suppl 1:S13-15.

Mihm S, Fayyazi A, and Ramadori G (1997): Hepatic expression of inducible nitric oxide synthase transcripts in chronic hepatitis C virus infection: relation to hepatic viral load and liver injury. Hepatology; 26(2):451-458.

Mohamed MK, (2004): Epidemiology of HCV in Egypt (2004): The Afro-Arab Liver Journal; vol 3, No2, pp 41-52.

Montgomery HAC, and Dymock, JF, (1961): The determination of nitrate in water. Analyst; 86, 414-416.

Moos V, Rudwaleit M, Herzog V, Höhlig K, Sieper J, and Müller B (2001): Association of genotypes affecting the expression of interleukin-1beta or interleukin-1 receptor antagonist with osteoarthritis. Erratum in: Arthritis Rheum; 44(7):1715.

Nemetz A, Nosti-Escanilla MP, Molnar T, Kope A, Kovacs A, Feher J, Tulassay Z, Nagy F, Garcia-Gonzalez MA, and Pena AS (1999): IL- 1B gene polymorphisms influence the course and severity of inflammatory bowel disease. Immunogenetics; 49: 527-531.

Paradis V, Mathurin P, Kollinger M, Imbert-Bismut F, Charlotte F, Piton A, Opolon P, Holstege A, Poynard T, and Bedossa P (1997): In situ detection of lipid peroxidation in chronic hepatitis C: correlation with pathological features. J Clin Pathol; 50(5):401-406.

Pârvu AE, Negrean V, Pleșca-Manea L, Cosma A, Drăghici A, Uifălean A, Moldovan R. (2005): Nitric oxide in patients with chronic liver diseases. Rom J Gastroenterol. Sep;14(3):225-230.

Pessayre D, Berson A, Fromenty B, and Mansouri A (2001): Mitochondria in steatohepatitis. Semin Liver Dis; 21(1):57-69.

Poynaru T, Ratziu V, Charlotte F, Goodman Z, McHutchison J, and Albrecht J (2001): Rates and risk factors of liver fibrosis progression in patients with chronic hepatitis C. J Hepatol; 34(5):730-739.

Sariçam T, Kircali B, and Köken T (2005): Assessment of lipid peroxidation and antioxidant capacity in non-alcoholic fatty liver disease. Turk J Gastroenterol; 16(2):65-70.

Seronello S, Sheikh MY, Choi J (2007): Redox regulation of hepatitis C in nonalcoholic and alcoholic liver; Free Radic Biol Med. Sep 15;43(6):869-882.

SPSS Inc. (1999):SPSS Base 10.0 for Windows User's Guide. SPSS Inc., Chicago IL.

Tanaka Y, Furuta T, Suzuki S, Orito E, Yeo AE, Hirashima N, Sugauchi F, Ueda R, and Mizokami M (2003): Impact of interleukin-1beta genetic polymorphisms on the development of hepatitis C virus-related hepatocellular carcinoma in Japan. J Infect Dis; 187(11):1822-1825.

Vendemiale G, Grattagliano I, Portincasa P, Serviddio G, Palasciamo G, and Altomare E (2001): Oxidative stress in symptom-free HCV carriers: relation with ALT flare-up. Eur J Clin Invest; 31(1):54-63.

Wang Y, Kato N, Hoshida Y, Yoshida H, Taniguchi H, Goto T, Moriyama M, Otsuka M, Shiina S, Shiratori Y, Ito Y, and Omata M (2003): Interleukin-Ibeta gene polymorphisms associated with hepatocellular carcinoma in hepatitis C virus infection. Hepatology; 37(1):65-71.

World Health Organization. Global surveillance and control of hepatitis C (1999): J Viral Hepat; 6:35-47.

Xu J, Cao H, Liu H, Wu ZY. Role of nitric oxide synthase and cyclooxygenase in hyperdynamic splanchnic circulation of portal hypertension (2008): Hepatobiliary Pancreat Dis Int; 7(5):503-508.

Zamora R, Vodovotz Y, and Billiar TR (2000): Inducible nitric oxide synthase and inflammatory diseases. Mol Med; 6(5):347-373.

Zhang PA, Li Y, Xu P, and Wu JM (2004): Polymorphisms of interleukin-1B and interleukin-1 receptor antagonist genes in patients with chronic hepatitis B. World J Gastroenterol; 10(12):1826-1829.

التعدد الشكلي لجين مضاد المستقبل انترايوكين-١ والأجهاد التأكسدي بين مرضي الإلتهاب الكبدي الوبائي

القيروسى سى .

عبد العزيز فتوح" - عفاف محمد السعيد" شيماء فتحي زكى"" - أحمد عبد السلام ستين"

* قسم الكرمياء - كلية العلوم - جامعة المنصورة - *وحده أمراض الوراثة -كلية الطب - جامعة المنصورة **مستشفي الأطفال الجامعي - كلية الطب - جامعة المنصورة - * كلية الطب - قسم الأطفال الجامعي المنصورة.

يعتبر جين الانترليوكين-١ أول السيتوكين إكتشافا حيث أنه له دور فعال في التحكم في الجهاز المناعي ضــد أي جسم غريب أو أي إصابة بعدوي فيروسية وكذلك في عمليات الأيض التي يقوم بها الكبد.

إن مرض الإلتهاب الكبدي الفيروسي الوبائي سي من أكثر الأمراض شيوعاً في محصر وأغلب دول العالم ويصاحب هذا المرض وجود تغيرات في جهد التأكسد بالجسم وذلك نتيجه وجود الفيروس وما يحصاحبه من التهابات وتلف في خلايا الكبد.

وفي هذه الدراسة تم إيجاد العلاقة بين التعدد الشكلي للجين الخاص بمضاد المستقبل انترليـوكين-١ ومـرض الإلتهاب الكبدي الوباني الفيروسي سي ومدي تأثيره على الجهد التأكسدي عند هولاء المرضي. فقد تـم عمـل دراسة للحمض النووي للجين الخاص بمضاد المستقبل انترليوكين-١ بأستخدام تفاعـل البـوليمريز المتسلـسل للمرضى المصابين باللإلتهاب الكبدي الفيروسي سي وعددهم ١١٠ شخص مـن المتـرددين علـي عيـادات الأمراض المتوطنة في مستشفى المنصورة الجامعي وايضا على حوالي ١٢٤ شخص من الأصـحاء الـذين لا يعانون من وجود أي أمراض وقد تم أخدهم من متبر عبين ببنك الدم بمستشفى الأطغال الجامعي وذلك للمقارنه.

ومن خلال هذه الدراسة فقد وجد أن للجين الخاص بمضاد المستقبل انترابوكين-١ العديد من الطرز الجيني بـين المرض المصابين بالإلتهاب الكبدي الوبائي الفيروسي سي وكذلك بين الأفراد الأصحاء. منها الطـراز الجينـي AIA2 وهو مميز في الأفراد الأصحاء عنه بين المصابين بالفيروس.أمـا الطـراز الجينـي IA1A وكـذلك A2A2 فان نسبة ظهوره في مرضى الإلتهاب الكبدي الفيروسي أعلى منها عند الأصحاء.

كما وجد أن أكسدة الدهون والمتمثلة في مركب المالونيل ثنائي الألدهيد فهو أعلى في المرضى عنه في الأصحاء وذلك لوجود فيروس سي وتأثيره على حالة الجهد التأكسدى في جسم المريض.أما المثق الحر اكسيد النتريك فإنه مميز في حالة الطراز الجينى A2A2 بين مرضى الإلتهاب الكبدي الوبائي الفيروسي سي.

ومن هنا نستنتج أن وجود فيروس سي قد أثر على الجهد التأكسدى وكذلك على الجهاز المنساعي وان الطسراز الجيني A2A2 مميز في مرضمي الإلتهاب الكبدي الوبائي الفيروسي سي وأن هنساك إحتماليسة للحمايسة مسن الإلتهاب الكبدي الوبائي الفيروسي سي في وجود الطراز الجيني A1A2.

