

MONITORING OF SOME SOIL MICROBES AND THEIR RELATIONSHIP TO SOIL SALINITY IN AL-HAMOUL REGION-KAFR EL-SHEIKH GOVERNORATE

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ABSTRACT

The soil samples were collected from the west to east side of the region. Most of the collected soil samples were highly saline, but salinity levels decreased as we headed east. The EC values ranged from 4.14 to 26.16. The viable counts of *R. leguminosarum* bv. *viceae*, *R. leguminosarum* bv. *trifolii* and *Azotobacter* varied through different sites, they ranged from log to log 0.84; log 2.29 to 0.84; log 2.23 to 4.26, respectively and the variations between readings were, sometimes, significant. But, there were no significant correlations between the estimated bacterial types and soil salinity. Total bacterial readings, however, showed high values, but, lowered in soil samples collected from the west sites. These bacterial counts were decreased as we headed east. They ranged from log 7.26 to log 9.46. However, there was no significant correlation between bacterial counts and soil salinity.

INTRODUCTION

Al-Hamool district located at North Delta region Kafr-El-Sheikh Governorate (31° 15' 30" N, 31° 9' 36" E), bounded from the North by Baltim and Burolos lake, from the South Bialla and Kafr-El-Sheikh Centers, from the East Bialla Center and from the West Kafr-El-Sheikh and Al-Riad Centers. The agricultural area was about 20000 feddan and called Al-Hamool due to large growth of *Cuscuta epithimum* parasite plant (Hamoul). The lands of this area are newly reclaimed, some of which have low or moderate productivity and others still under reclamation. Its soils are saline and/or alkaline with varied degrees. The most problems of this area are salinity, alkalinity, low soil fertility and scarcity of low quality of irrigation water as this area is located at the end of irrigation and drainage canals. The fields have been irrigated with mixed polluted water composed from clean (Tera canal) and drainage water; sewage, industrial and agricultural (Ketchener drainage canal) with ratio of 1:1. This water is of low quality. Some of the fields irrigated totally with drainage water and others do not get enough water and suffer from drought.

A number of bacterial species belonging to genera *Azospirillum*, *Alcaligenes*, *Arthrobacter*, *Acinetobacter*, *Bacillus*, *Burkholderia*, *Enterobacter*, *Erwinia*, *Flavobacterium*, *Pseudomonas*, *Rhizobium* and *Serratia* are associated with the plant rhizosphere and are able to exert a beneficial effect on plant growth (Egamberdiyeva, 2000 and Tilak *et al.*, 2000). Utilization of plant growth promoting rhizobacteria (PGPR) in order to increase the productivity may be a viable alternative to organic fertilizers which also helps in reducing the pollution and preserving the environment in the spirit of an ecological agriculture (Stefan *et al.*, 2008). Thus rhizospheric bacteria can be a promising source for plant growth promoting agent in agriculture and are commonly used as inoculants for improving the growth and yield of agricultural

crops (Chaiharin *et al.*, 2008). Biological nitrogen fixation contributes 180×10^7 metric tons/year globally, out of which symbiotic associations' produces 80% and the rest comes from free-living or associative systems (Graham, 1998). These include symbiotic nitrogen fixing (N_2 -fixing) forms, viz. *Rhizobium*, the obligate symbionts in leguminous plants and Frankia in non-leguminous trees, and non-symbiotic (free-living, associative or endophytic) N_2 -fixing forms such as cyanobacteria, *Azospirillum* and *Azotobacter*.

Agricultural crops are exposed to many stresses that are induced by both biotic and abiotic factors. These stresses decrease yields of crops and represent barriers to the introduction of crop plants into areas that are not suitable for crop cultivation. The occurrence and activity of soil microorganisms are affected by a variety of environmental factors as well as plant-related factors (species, age). Abiotic stress factors include high and low temperature, salinity, drought, flooding, ultraviolet light, air pollution (ozone) and heavy metals. The yield losses associated with abiotic stresses can reach 50% to 80%, depending on the crop. In many semi-arid and arid regions of the world, crop yield is limited due to increasing salinity of irrigation water as well as soil salinity. The inoculation of salt-stressed plants with PGPR strains alleviates the salinity stress in plants. Soil salinity is one of the most severe factors limiting nodulation, yield and physiological response in soybean. An increase in salinity in the soil causes a physiological response or disorder in lettuce plants (Han and Lee, 2005). The long-term goal of improving plant-microbe interactions for salinity affected fields and crop productivity can be met with an understanding of the mechanism of osmoadaptation in *Azospirillum* sp. The synthesis and activity of nitrogenases in *A. brasilense* is inhibited by salinity stress (Tripathi *et al.*, 2002). They reported that in *Azospirillum* sp. there is an accumulation of compatible solutes such as glutamate, proline, glycine betaine and trehalose in response to salinity/osmolarity; proline plays a major role in osmoadaptation through increase in osmotic stress that shifts the dominant osmolyte from glutamate to proline in *A. brasilense*. *Azospirillum*-inoculated sorghum plants had more water content, higher water potential, and lower canopy temperature in their foliage. Hence, they were less drought-stressed than noninoculated plants (Saleena *et al.*, 2002). The PGPR containing ACC deaminase (1-aminocyclopropane-1-carboxylate) are present in various soils and offer promise as a bacterial inoculum for improvement of plant growth, particularly under unfavourable environmental conditions such as flooding, heavy metals, phytopathogens, drought and high salt. Ethylene is an important phytohormone, but over-produced ethylene under stressful conditions can result in the inhibition of plant growth or death, especially for seedlings. PGPR containing ACC deaminase can hydrolyze ACC, the immediate precursor of ethylene, to F-ketobutarate and ammonia, and in this way promote plant growth. Inoculation of crops with ACC deaminase-containing PGPR may assist plant growth by alleviating deleterious effects of salt stress ethylene (Belimov *et al.*, 2001).

Microbial density in soil usually are correlated with organic matter percentage and sometimes biological inoculants such as N_2 -fixing and phosphate-dissolving bacteria (Nour El-Din and Talha, 2011). They found, in

long term experiment, that compost addition and inoculation with N₂-fixing and phosphate-dissolving bacteria enhanced soil organic matter percentage and available N,P and K contents. Miransari (2010), also claimed that biological fertilization caused increase of N and P as well as microelements in soil. On the other hand, there are some factors like salinity and heavy metals pollution negatively affecting microbial density of soil which may reflected on soil organic matter and nutrient contents.

The aim of the present investigation is the monitoring of viable counts of total bacteria, *R. leguminosarum* bv. *viceae*, *R. leguminosarum* bv. *trifolii* and *Azotobacter* in rhizosphere soil of Al-Hamoul Center, Kafr-El-Sheikh Governorate and their relationship with salinity level of soil, in order to give us an indicator for the soil quality and fertility.

MATERIALS AND METHODS

Materials:

Microbial media:

Medium 1 for counting of *Azotobacter*: Vancura and Mucura (1960), it composed of sucrose, 30 g; K₂HPO₄, 0.16 g; NaCl, 0.2 g; MgSO₄·7H₂O, 0.2 g; CaCO₃, 2.0 g; FeSO₄, 0.005 g; Na₂MO₄, 0.005 g; NaBO₂, 0.005 g and distilled water, 1 liter.

Medium 2: for total counting of bacteria (Allen, 1909)

Glucose, 10.0 g; K₂HPO₄, 1.0 g; soil extract, 100 ml; distilled water, 900 ml; agar, 10 g; pH adjusted at 6.8-7.0. Autoclaved at 121°C for 10 min.

Soil extract:

Soil extract is prepared by heating 1000 g of grinding soil with 1000 ml of tap water in the autoclave for 30 minutes. A small amount of calcium carbonate (2.0 g/l) is added on the soil suspension and filtered through a double paper filter. The filtrate was divided into quantities each equal 100 ml and then sterilized.

Methodes:

Site of samples: soil and plant (faba bean and Egyptian clover) samples sites were determined using the detailed spatial map of Al-Hamool district, as the samples designed every 2 km with the rate of one composite sample per 200 km².

Soil sampling: Using spade, the soil bulk around the plant (30 cm depth) was removed and backed in plastic bags. Three faba bean samples were taken from each field and other three samples of *Trifolium Alexandrinum* were bring as possible from the same site. Different microbial groups were counted (total count of bacteria, *Azotobacter* *R. leguminosarum* biovar *viceae* and *R. leguminosarum* biovar *trifolii*).

***R. leguminosarum* biovar *viceae* and *R. leguminosarum* biovar *trifolii* count in soil:** The counting was done using MPN technique reported by Somasegaran and Hoben (1980).

***Azotobacter* count:** MPN method, using medium of Vancura and Mucura (1960) was performed according to Allen (1909).

Total count of bacteria:

Soil extract solidified medium was poured in Petri dishes, then inoculated with the appropriate dilutions of the soil samples and incubated at 28°C for three days. The appeared colonies were counted and the count per 1 g air dry soil was calculate as reported by Allen, (1969).

EC: EC of soils were estimated according to Richards (1964).

Statistical analysis:

Data obtained were subjected to the analysis of variance and treatment means were compared using the L.S.D methods according to Steel and Torrie (1980).

RESULTS AND DISCUSSION

Through monitoring EC values (dS/m) for soils of many villages of Al-Hamoul Center, Kafr El-Sheikh Governorate (Table 1), it is noted that most of soil samples brought about from the Eastern villages recorded higher EC values (the first 20 sample), and their EC values ranged from 4,40 to 12,71 dS/m. However, the EC values of the collected samples lowered with the direction of west, the EC values of the following twenty western sites ranged from 4,30 to 10,60 dS/m except for two sites (No. 20 and 26) which were estimated as 26,16 and 22,60 dS/m respectively. The following last twenty samples from the western direction recorded the lowest EC values, ranged from 4,16 to 7,11 dS/m except the sites No. 02 and 03 which was of relatively high records (9,7 and 10,32 dS/m).

The Al-Hamoul region consider from the sites under reclamation in Kafr El-Sheikh Governorate. Soil of some locations were reclaimed and others still had high salinity nature. The west site is the nearest from Al-bourollos Lake and most of its fields used as fish cultures, and others recently used as agricultural fields. Therefore, salinity levels in this fields still high. With the progress toward east, the soils have been reclaimed and their EC values lowered except some fields that may be neglected by farmers. The main difficulty of reclamation of these soils is the use of drainage water in irrigation (Zein et al., 2002).

The results of monitoring *R. leguminosarum* biovar *viceae* and *R. leguminosarum* biovar *trifolii* of Al-Hamoul soils relatively varied, and there are significant variations between most of these records (Table 2 and 3). However, there was no significant correlation between salinity levels of soil samples and log number of these microbes (Fig 1). The lowest monitored *R. leguminosarum* biovar *viceae* count was log 4.00 soil sample No. 49, while, the highest count recorded log 6,84 which repeated frequently at many sites of the studied area. On the other hand, the lowest number of *R. leguminosarum* biovar *trifolii* (log 2.39) was recorded for soil sample No. 24 and the highest value were 6,84 which recorded at many sites of the studied area.

The high counts of native *R. leguminosarum* biovar *viceae* and *R. leguminosarum* biovar *trifolii* may be attributed to continuous cultivation of faba bean and Egyptian berseem since long period, therefore the indigenous specific rhizobia spread and localized in soils of the region.

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The macro-symbiont plants brought from the region were well nodulated (not presented data) which explains the potentiality of these rhizobia for root infection and formation of nodules. Nour El-Din (1997) reported that the fields had been cultivated with *Trifolium alexandrinum* or *Vicia faba* had high density of the specific native rhizobia. But, in the present study there were no significant correlation between the counts of the microsymbionts in the plants rhizosphere and the salinity levels of the soil samples. This may be attributed to two reasons: the first is the formation of mutations in the native rhizobial populations lead to presence of new salt tolerant strains potentially able to live effectively under these stress conditions (Zahran, 1999). The second reason is that fields of this region had been irrigated with sewage water which have a lot of nutrients and organic matter, which enriched the soil and increased the soil water holding capacity (Mohammed, 2011). Wei *et al.* (1980) claimed that sewage sludge application to agricultural soils is an economical way of disposal. It improves the physical characteristics of the soil and increases organic matter content and essential plant nutrients, in particular N and P (Ferreira *et al.*, 1990). Sewage sludge contains numerous components required for microbial growth and may increase the activity of soil microorganisms (Speaker *et al.*, 1988), including rhizobial growth. But, the heavy amounts of sewage amendments for long times may negatively affect N₂-fixers efficiency which may attributed to high amounts of heavy metals (Chaudhary *et al.*, 2004) and increase amounts of nitrates (Obbard *et al.*, 1993).

Azotobacter viable counts were more less numbers per gm of rhizosphere soil than *R. leguminosarum* biovar *viceae* and *R. leguminosarum* biovar *trifolii*, the counts ranged from log 2 to log 4.26 (Table, 4). As found for counts of *R. leguminosarum* biovar *viceae* and *R. leguminosarum* biovar *trifolii*, there was no positive correlation between the *Azotobacter* counts and salinity degrees (Fig, 1). In case of total bacteria, the counts increased as we head east of the province (Table 5), whereas, the first 20 samples brought about from the far west of the province (Al-Hamoul Center) mostly gave relatively low counts (less than log 4), while most of the following 20 samples toward east, recorded counts more than log 4, but, the following 20 samples toward east (from sample No. 41 to No. 60) attained the highest numbers (mostly higher than log 4.7) and there were significant variation between most samples. However, there was no significant correlation between the bacterial counts and soil salinity levels. The lower viable counts of *Azotobacter* than rhizobial counts may be due to the direct contact of *Azotobacter* cells with salty and heavily polluted soils with heavy metals because of the irrigation with sewage water (Zein *et al.*, 2002). Rhizobia harbored within nodules gave them relative protection against salinity and heavy metals. On the other hand, extensive planting of the host macrosymbiont plants resulted in increase of the specific rhizobia in the soil. Total bacterial counts noted to be lower in soil of west sites which were close to Al-Brolas lake, and had no enough reclamation, therefore the soils still highly saline as shown in Table 1. The deleterious effect may result from sensitivity of some sensitive bacterial types to unfavorable conditions which may lead to decrease of total count.

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The total bacterial counts increased as we head east, this may be due to increase of reclamation efforts as we head east of the region.

CONCLUSION

Al-Hamoul region is of the locations recently reclaimed, thus, there were variations in its soils fertility. Monitoring of soil salinity and microbial biomass of these soils are of important which give us an indicator for soil quality and fertility. The survey indicated that most of west sites are highly in salin than the other sites of the region. The *R. leguminosarum* bv. *viceae*, *R. leguminosarum* bv. *trifolii* and *Azotobacter* were tolerant to salinity and other unfavorable environmental conditions. The total bacterial counts mostly varied. This area needs clean irrigation water and more effective reclamation processes to increase the economic value of unit area and improve living level of the inhabitants.

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رصد أعداد ميكروبات التربة المثبتة للآزوت الجوي وعلاقتها ببعض خصائص
التربة بمنطقة الحامول – محافظة كفرالشيخ
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مركز البحوث الزراعية-معهد بحوث الاراضي و المياه والبيئة-الجيزة-القاهرة

يقع مركز الحامول بالمنطقة الشمالية الغربية من محافظة كفرالشيخ، وتبلغ مساحة الاراضي الصالحة للزراعة بالمركز حوالي ٢٠٠٠٠٠ فدان، وهذه الاراضي حديثة الاستصلاح، فمنها من تم استصلاحه والاخر مازال تحت الاستصلاح. وتقع أراضي مركز الحامول في نهاية الترع والمصارف، لذلك فانها تعاني من ندرة مياه الري النظيفة، وتروي هذه الاراضي اما بمياه الصرف (صحي وزراعي وصناعي) أو بالمياه المخلوطة من مياه ترعة بحر تيرة النظيفة ومياه مصرف كتشنر الملوثة والغير معالجة، مما يجعل الاراضي بهذه المنطقة تعاني من الملوحة والتلوث بالعناصر الثقيلة وكذلك الميكروبات الممرضة. لذلك فاننا نهدف في هذه الدراسة الي رصد لحالة التربة بهذه المنطقة من ناحية درجات ملوحتها واعداد بعض الميكروبات المهمة بمنطقة جذور نباتي البرسيم المسقاي و الفول البلدي، كي تعطينا مؤشرا عن حالة هذه الاراضي و مدي خصوبتها. كانت معظم العينات المجلوبة من مركز الحامول عالية الملوحة، وكانت ملوحة الاراضي تقل كلما اتجهنا ناحية الشرق، وقد وجد تباين في أعداد ميكروبات الازوتوباكتر و ريزوبيا الفول البلدي و ريزوبيا البرسيم المسقاي من موقع الي اخر. وعلي الرغم من وجود هذه الاعداد بوفرة، فلم يظهر ارتباط معنوي بين هذه الاعداد و قيم ملوحة التربة . كانت اعداد البكتريا الكلية بالتربة كبيرة نسبيا، ولكنها كانت أقل كلما تجهنا ناحية الغرب. أيضا لم يوجد ارتباطا معنويا بين الاعداد الكلية للبكتيريا بالتربة و قيم الملوحة لها. لذلك فاننا نوصي بتوصيل مياه الري النظيفة لهذه المنطقة للتقليل من ملوحة الاراضي و زيادة انتاجيتها و كذلك تقليل تلوث الاراضي والمزروعات بالعناصر الثقيلة والميكروبات الممرضة بهدف حماية الصحة العامة زيادة انتاجية الفدان لهذه المنطقة المنكوبة.

قام بتحكيم البحث

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Table 1: Monitoring of soil samples EC, dS/m, (soil extract) collected from Al-Hamoul region.

Sample No.	Coordination		EC	Sample No.	Coordination		EC	Sample No.	Coordination		EC
	X	Y			X	Y			X	Y	
1	-	-	10.81b-h	21	36316881	3477281	7.40f-p	41	36317113	3473392 M	4.16p
2	-	-	10.70b-i	22	36310996	3479831	10.27b-l	42	36317490	3474084 M	3.91p
3	-	-	13.03bc	23	36313874	3480376	11.11b-g	43	36317600	3470284 M	4.23op
4	-	-	12.07b-e	24	36313078	3481430	7.78c-p	44	36317748	3470908 M	0.22i-p
5	-	-	9.27b-p	25	36313730	3480878	7.79e-p	45	36317871	3478703 M	4.30p
6	-	-	8.89b-p	26	36314878	3480837	7.03f-p	46	36317113	3480170 M	4.16k-p
7	-	-	11.48b-f	27	36317678	3477242	7.97e-p	47	36317019	3481278 M	4.08m-p
8	-	-	11.00b-f	28	36317402	3473880	0.76g-p	48	36310937	3482127 M	4.92k-p
9	-	-	11.26b	29	36321940	3473489	0.73g-p	49	36310374	3483449 M	0.00k-p
10	36314240	3476498	13.70bc	30	36320774	3477907-M	4.93k-p	50	36317741	3482830 M	7.47f-p
11	36310002	3476081	7.43d-p	31	36318913	3477900-M	4.80 l-p	51	36317609	3482824 M	7.11e-p
12	36310012	34774	7.40d-p	32	36318078	3470039-M	4.96k-p	52	36318104	3484076 M	9.70b-o
13	36310000	3478477	8.42b-p	33	36317341	3471000	4.30nop	53	36318188	3487271 M	10.22b-k
14	36314702	3478121	12.03b-f	34	36317284	3473841-M	10.70b-m	54	36318138	3487270 M	7.22f-p
15	36314220	3476027	13.36b	35	36317700	3478831-M	26.17a	55	36319642	3489192 M	4.96k-p
16	36314270	3479703	12.76bcd	36	36310077	3477322-M	23.70a	56	36322277	3470190 M	4.70m-p
17	36310076	3472270	12.71bcd	37	36317411	3481142-M	9.79b-n	57	36322180	3470190 M	4.70m-p
18	36307098	3472499	8.06b-p	38	36311349	3477774 M	7.03f-p	58	36321179	3473170 M	4.97k-p
19	36317091	3473117	7.70d-p	39	36312039	3477711 M	0.30j-p	59	36321847	3470740 M	0.42h-p
20	36317108	3474928	7.14e-p	40	36313704	3470022 M	0.19j-p	60	36321773	3471802 M	4.76 l-p

Means with different letters differ significantly at $P < 0.05$.
 Coordination: is the reading of X and Y taking by GPS device.

Table 2: Monitoring number (log) of soil samples *R. leguminosarum* biovar *viceae* (R.V.) collected from Al-Hamoul region.

Sample No.	Coordination		R.V.	Sample No.	Coordination		R.V.	Sample No.	Coordination		R.V.
	X	Y			X	Y			X	Y	
1	-	-	o,31c-f	21	36317881	3477281	o,8εabc	ε1	36317713	3473392 VM	o,0yc-g
2	-	-	o,31c-f	22	36310996	3479831	o,8εabc	ε2	36317490	34740.8ε .M	o,0yc-g
3	-	-	o,8εabc	23	36313874	3480036	o,8εabc	ε3	3631760.	347028ε 1M	o,0εc-g
4	-	-	o,8εabc	24	36313.78	348143.	o,70bcd	ε4	36317248	34709.8 3M	o,00b-e
5	-	-	o,8εabc	25	36313730	3480878ε	o,00b-e	ε5	36317871	34787.2 1M	o,0yc-g
6	-	-	o,8εabc	26	36314878	3480837ε	o,00b-e	ε6	3631713.	3480170 .M	o,0yc-g
7	-	-	o,31c-f	27	36317678	3472242	ε,70d-g	ε7	36317.19	3481278 .M	o,27c-g
8	-	-	o,31c-f	28	36317402	3473880.	ε,23fg	ε8	36310937	3482127 .M	ε,01efg
9	-	-	o,31c-f	29	3632194.	3473489	o,8εabc	ε9	36310374	3483449 .M	ε,0.g
10	3631424.	3631424.	o,31c-f	30	3632.274	34779.7-2M	o,8εabc	00	36317241	348283. .M	o,0εc-g
11	363100.2	363100.2	o,8εabc	31	36318913	3477900-2M	o,8εabc	01	36317609	348282ε 1εM	o,8εabc
12	36310012	36310012	o,8εabc	32	36318.78	3470.29-.M	o,8εabc	02	363181.ε	3484077 .M	o,8εabc
13	3631000.	3631000.	o,8εabc	33	36317341	34710.0	o,00b-e	03	36318188	3487271 .M	o,8εabc
14	363146.2	363146.2	o,0εc-f	34	36317284	347384ε1-1M	o,27c-f	04	36318138	348727. .M	o,8εabc
15	3631422.	3631422.	ε,0εc-g	35	363177.0	3477831-9M	o,00b-e	05	36319642	3489192 2M	o,8εabc
16	36314270	36314270	ε,82c-g	36	36310077	3477322-2M	o,8εabc	06	36322277	3470.190 9M	o,8εabc
17	36310076	36310076	o,31c-f	37	36317411	3481142-1M	o,0εc-g	07	3632218.	3470.190 .M	o,8εabc
18	36307098	36307098	o,0εc-g	38	36311349	3477774ε .M	o,0εc-g	08	36321179	347317. .M	o,8εabc
19	36317.91	36317.91	ε,23g	39	36312039	3477711 2M	o,0εc-g	09	36321847	3470.7ε. .M	o,8εabc
20	36317108	36317108	o,0εc-g	40	363137.ε	3470022 .M	ε,77d-g	10	36321773	34718.2 2M	o,8εabc

Means with different letters differ significantly at P < 0.05.

Coordination: is the reading of X and Y taking by GPS device.

Table 3: Monitoring number (log) of soil samples *R. leguminosarum* biovar *trifolii* (R.T.) collected from Al-Hamoul region.

Sample No.	Coordination		R.T.	Sample No.	Coordination		R.T.	Sample No.	Coordination		R.T.
	X	Y			X	Y			X	Y	
1	-	-	0,31abc	21	37317881	3577281	0,0εa-f	ε1	37317713	3573392 VM	0,0γc-g
2	-	-	0,8εa	22	37310996	3579831	ε,77b-g	ε2	37317490	357408ε .M	0,0γc-g
3	-	-	0,8εa	23	3731387ε	3ε8.037	ε,77b-g	ε3	3731760.	357028ε 1M	0,0εc-g
ε	-	-	0,8εa	2ε	37313078	3ε81ε30.	2,39b-g	εε	373172ε8	3570908 2M	0,00b-e
0	-	-	0,00ab	20	37313730	3ε0878ε	3,08hij	ε0	37317871	3578702 1M	0,0γc-g
6	-	-	0,38abc	26	3731ε878	3ε0837ε	ε,0.c-h	ε6	37317130.	3ε80170 .M	0,0γc-g
7	-	-	ε,77b-g	27	37317678	3ε722ε2	ε,23f-i	ε7	37317019	3ε81278 .M	0,27c-g
8	-	-	0,31abc	28	37317ε02	3ε73880.	3,0.ij	ε8	37310937	3ε82127 .M	ε,01efg
9	-	-	0,31abc	29	373219ε0.	3ε73ε89	ε,3.d-i	ε9	3731037ε	3ε83εε9 .M	ε,0.g
10	3731ε2ε0.	3731ε2ε0.	0,31abc	30	3732027ε	3ε77907.2M	0,8εa	00	373172ε1	3ε82830 .M	0,0εc-g
11	37310002	37310002	0,31abc	31	37318913	3ε779000.2M	0,8εa	01	37317609	3ε8282ε 1εM	0,8εabc
12	37310012	37310012	0,31abc	32	37318078	3ε70039-.M	0,31abc	02	3731810ε	3ε8ε077 .M	0,8εabc
13	37310000.	37310000.	0,8εa	33	373173ε1	3ε71000	ε,27e-j	03	37318188	3ε87271 .M	0,8εabc
1ε	3731ε702	3731ε702	ε,80b-g	3ε	3731728ε	3ε738ε1-1M	ε,0.g-j	0ε	37318138	3ε87270 .M	0,8εabc
10	3731ε220.	3731ε220.	ε,30d-i	30	37317700	3ε77831-9M	0,0εa-f	00	373197ε2	3ε09192 2M	0,8εabc
16	3731ε270	3731ε270	ε,30d-i	36	37310077	3ε77322-2M	0,8εa	06	37322277	3ε70190 9M	0,8εabc
17	37310076	37310076	ε,00g-j	37	37317ε11	3ε811ε2-1M	0,31abc	07	37322180.	3ε70190 .M	0,8εabc
18	37307098	37307098	ε,23f-i	38	373113ε9	3ε7777ε .M	ε,77b-g	08	37321179	3ε73170 .M	0,8εabc
19	37317091	37317091	ε,23f-i	39	37312039	3ε77711 2M	ε,0.c-h	09	373218ε7	3ε707ε0 .M	0,8εabc
20	37317108	37317108	0,0εa-f	ε0	3731370ε	3ε70022 .M	ε,0.c-h	70	37321773	3ε71802 3M	0,8εabc

Means with different letters differ significantly at $P < 0.05$.
 Coordination: is the reading of X and Y taking by GPS device.

Table 4: Monitoring number (log) of soil samples *Azotobacter* (Az.) collected from Al-Hamoul region.

Sample No.	Coordination		Az.	Sample No.	Coordination		Az.	Sample No.	Coordination		Az.
	X	Y			X	Y			X	Y	
1	-	-	3,00C-f	21	37317881	3477281	ε,27a	ε1	37317713	3473392 VM	2,77
2	-	-	3,00C-f	22	37310997	3479831	ε,00a	ε2	37317490	3474084 M	2,77
3	-	-	3,00abc	23	37313874	3480037	3,00C-f	ε3	37317700	3470284 VM	2,00
4	-	-	3,00abc	24	37313078	3481430	2,27ef	ε4	37317248	3470908 VM	2,00
5	-	-	3,27b-e	25	37313730	3481784	3,27b-e	ε5	37317811	3478702 VM	2,27
6	-	-	3,00C-f	26	37314878	3482374	3,00C-f	ε6	37317130	3480170 M	2,77
7	-	-	2,77C-f	27	37317778	3482242	2,00C-f	ε7	37317019	3481278 M	2,00
8	-	-	2,70C-f	28	37317402	3483880	3,00C-f	ε8	37310937	3482127 M	2,73
9	-	-	2,00C-f	29	37321940	3483489	3,00C-f	ε9	37310374	3483449 M	3,23
10	37314240	37314240	2,73C-f	30	37320274	3487907-2M	2,77C-f	50	37317241	3482830 M	3,00
11	37310002	37310002	3,00C-f	31	37318913	3487900-2M	3,27ef	51	37317609	3482824 VM	2,77
12	37310012	37310012	2,77C-f	32	37318078	3480039-M	2,77C-f	52	37318104	3484077 M	2,27
13	37310000	37310000	2,00C-f	33	37317341	3481000	3,27b-e	53	37318188	3487221 M	2,27
14	37314602	37314602	2,27ef	34	37317284	3483841-1M	2,77C-f	54	37318138	3487220 M	2,77
15	37314220	37314220	2,73C-f	35	37317700	3487831-9M	2,27ef	55	37319742	3489192 VM	3,00
16	37314270	37314270	2,73C-f	36	37310077	3487332-2M	3,27b-e	56	37322277	3480190 VM	3,23
17	37310076	37310076	2,27ef	37	37317411	3481142-1M	3,00abc	57	37322180	3480190 M	3,23
18	37307098	37307098	2,00C-f	38	37311349	3487774 M	2,27ef	58	37321179	3483170 M	3,00
19	37317091	37317091	3,00-f	39	37312039	3487711 VM	2,27ef	59	37321847	3480740 M	2,77
20	37317108	37317108	3,00C-f	40	37313704	3480022 M	2,00ef	60	37321773	3481802 VM	2,00

Means with different letters differ significantly at $P < 0.05$.

Coordination: is the reading of X and Y taking by GPS device.

Table 2: Monitoring number (log) of soil samples total bacteria (T.B.) collected from Al-Hamoul region.

Sample No.	Coordination		T.B.	Sample No.	Coordination		T.B.	Sample No.	Coordination		T.B.
	X	Y			X	Y			X	Y	
1	-	-	4,0Ypqr	21	37317881	3477281	8,0Ym-q	41	3731713	3473392 M	9,1Ya-g
2	-	-	4,0Ypqr	22	37310996	3479831	8,0Yn-q	42	37317490	3474084 M	9,2Ya-e
3	-	-	8,0Ym-q	23	37313874	3480536	8,1Y l-p	43	3731760	3470284 M	9,3Yabc
4	-	-	8,2Y l-p	24	37313078	3481430	8,1Y l-	44	37317248	3470908 M	9,1Ya-f
5	-	-	8,1Y l-p	25	37313730	3480878	8,1Y l-p	45	37317871	3478703 M	8,9Yb-h
6	-	-	4,3Ypqr	26	37314878	3480374	8,0Y n-q	46	3731713	3480170 M	9,1a-h
7	-	-	8,1Y l-p	27	37317678	3472242	8,10 l-p	47	37317019	3481278 M	9,08a
8	-	-	8,3Yi-n	28	37317402	3473880	8,2Yk-p	48	37310937	3482127 M	9,3Yabc
9	-	-	4,0Yqr	29	37321940	3473489	8,1Y l-p	49	37310374	3483449 M	8,81d-j
10	37314240	37314240	4,3Yr	30	37320274	3477906 M	8,3Yj-o	50	37317241	3482830 M	8,2Y l-p
11	37310002	37310002	4,810-r	31	37318913	3477900 M	8,3Yj-o	51	37317609	3482824 M	8,2Yk-p
12	37310012	37310012	4,8Ypqr	32	37318078	3470039 M	8,1Y l-p	52	37318104	3484066 M	8,8Yd-j
13	37310000	37310000	4,8Ypqr	33	37317341	3471000	8,2Yf-l	53	37318188	3487221 M	8,8Yd-j
14	37314602	37314602	4,7Ypqr	34	37317284	3473841 M	8,3Y i-n	54	37318138	3487220 M	8,8Yc-i
15	37314220	37314220	4,7Ypqr	35	37317700	3477831 M	8,2Yg-l	55	37319642	3489192 M	8,8Yd-j
16	37314270	37314270	4,9Yn-q	36	37310077	3477332 M	9,4Yab	56	37322277	3470190 M	8,8Yd-j
17	37310076	37310076	4,8Yn-r	37	37317411	3481142 M	9,3Yab	57	37322180	3470190 M	8,8Yd-j
18	37307098	37307098	4,8Ypqr	38	37311349	3477764 M	9,3Yabc	58	37322119	3473170 M	8,08h-m
19	37317091	37317091	4,8Ypqr	39	37312039	3477711 M	9,4Ya	59	37321847	3470740 M	8,8Ye-k
20	37317108	37317108	4,9Yn-q	40	37313704	3470022 M	9,2Yab	60	37321773	3471802 M	8,8Yd-j

Means with different letters differ significantly at $P < 0.05$.

Coordination: is the reading of X and Y taking by GPS device.

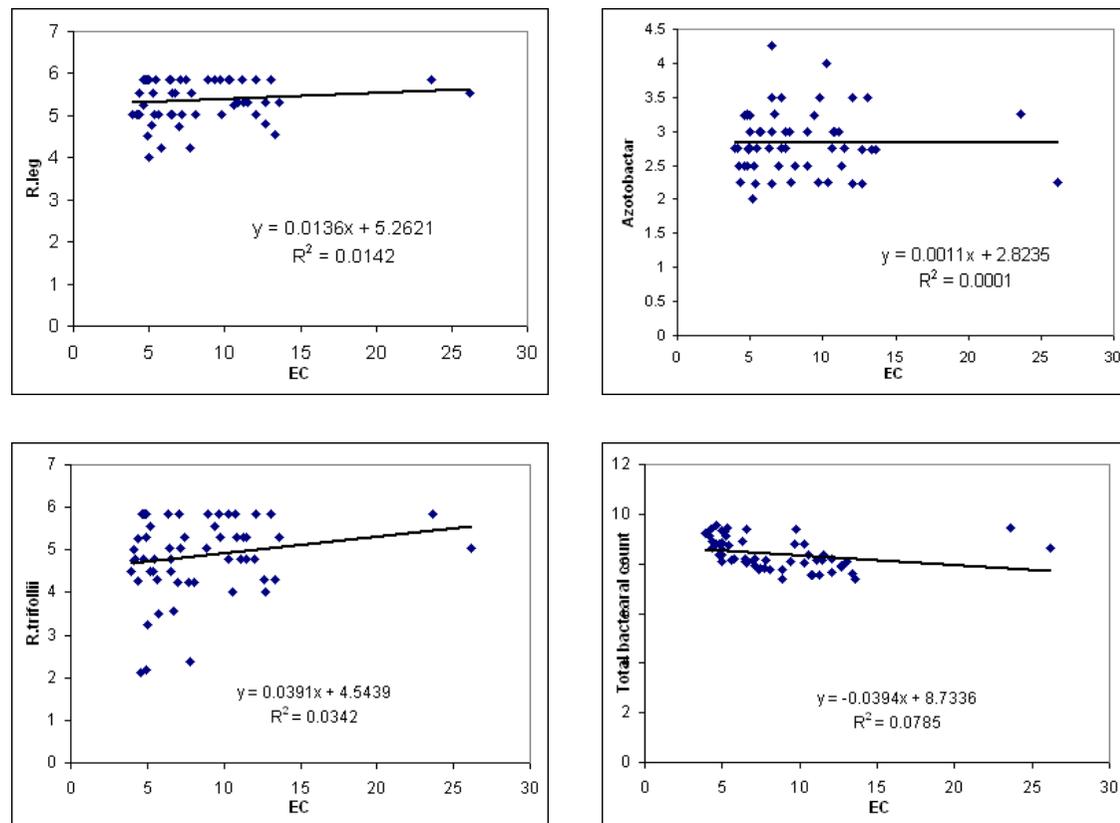


Figure 1: Correlations between salinity and log numbers of different studied microbes.